



The Sublethal Effects of the Entomopathic Fungus *Leptolegnia chapmanii* on Some Biological Parameters of the Dengue Vector *Aedes aegypti*

Authors: Pelizza, S.A., Scorsetti, A.C., and Tranchida, M.C.

Source: Journal of Insect Science, 13(22) : 1-8

Published By: Entomological Society of America

URL: <https://doi.org/10.1673/031.013.2201>



The sublethal effects of the entomopathic fungus *Leptolegnia chapmanii* on some biological parameters of the dengue vector *Aedes aegypti*

S.A. Pelizza^{1,2a*}, A.C. Scorsetti^{2b}, M.C. Tranchida^{2c}

¹CEPAVE (Centro de Estudios Parasitológicos y de Vectores) CCT-La Plata-CONICET-UNLP, La Plata (1900) Argentina

²Instituto de Botánica Carlos Spegazzini, Facultad de Ciencias Naturales y Museo, Universidad Nacional de La Plata (1900), Argentina

Abstract

The mosquito *Aedes aegypti* (L.) (Diptera: Culicidae) is the primary vector of dengue in the Americas. The use of chemical insecticides is recommended during outbreaks of dengue in order to reduce the number of adult mosquitoes; however, because *Ae. aegypti* is highly synanthropic, the use of insecticides in densely populated areas is a dangerous practice. *Leptolegnia chapmanii* Seymour (Straminipila: Peronosporomycetes) is an entomopathogenic microorganism that has demonstrated marked pathogenicity toward the larvae of a number of mosquito species, with little or no effect on non-target insects. Therefore, the purpose of this study was to determine the sublethal effects of *L. chapmanii* on fecundity, number of gonotrophic cycles, fertility, and relationship between wing length and fecundity in *Ae. aegypti* females. *Ae. aegypti* females that survived infection with *L. chapmanii* laid fewer eggs, had a smaller number of gonotrophic cycles, had shorter wings, and were less fertile than controls. This is the first study on the sublethal effects experienced by specimens of *Ae. aegypti* that survived infection with zoospores of *L. chapmanii*. Although field studies should be carried out, the results obtained in this study are encouraging because the high and rapid larval mortality caused by *L. chapmanii* coupled with the reduction of reproductive capacity in *Ae. aegypti* females seem to cause a significant reduction in the number of adults in the mid and long term, thereby reducing the health risks associated with *Ae. aegypti*.

Keywords: biocontrol, dengue, fecundity, fertility, gonotrophic cycles, mosquitoes, zoospores

Correspondence: ^a sebastianpelizza@conicet.gov.ar, ^b ascorsetti@conicet.gov.ar, ^c ctranchida@conicet.gov.ar,

*Corresponding author

Editor: Michael Adams was editor of this paper.

Received: 23 November 2011 **Accepted:** 9 October 2012

Copyright: This is an open access paper. We use the Creative Commons Attribution 3.0 license that permits unrestricted use, provided that the paper is properly attributed.

ISSN: 1536-2442 | Vol. 13, Number 22

Cite this paper as:

Pelizza SA, Scorsetti AC, Tranchida MC. 2013. The sublethal effects of the entomopathic fungus *Leptolegnia chapmanii* on some biological parameters of the dengue vector *Aedes aegypti*. *Journal of Insect Science* 13:22. Available online: <http://www.insectscience.org/13.22>

Introduction

The mosquito *Aedes aegypti* (L.) (Diptera: Culicidae), widely distributed in tropical and subtropical regions, is highly adapted to urban environments. It is frequently found inside or near houses and plays an important role in the transmission of arboviruses such as dengue and urban yellow fever (Belinato et al. 2009).

Currently, control of *Ae. aegypti* is attained mainly with chemical larvicides that target the insect's central nervous system. However, the long history of insecticide use has led to the development of resistant populations all over the world. Because of this resistance, the study of novel tools to control *Ae. aegypti* and other insects of medical importance is a major area of interest (Zaim and Guillet 2002).

The use of natural enemies to control mosquitoes, based mainly on the use of products based on *Bacillus thuringiensis* var. *israelensis* (Bti), has a successful track record (Ochoa et al. 2009). Formulations of Bti are effective against larvae of *Ae. aegypti*, but its use is limited because of its high cost and low residual power, requiring regular applications of the product with the consequent increase in the cost of control programs.

Several chemical larvicides and mosquito control agents have been shown to manifest delayed effects at sublethal doses in surviving mosquitos (Simsek et al. 2009). In laboratory studies, Aduelo-Silva and Spielman (1984) have shown that inefficient larvicide reduces larval competition among survivors and increases the density and average body size of the resulting adult population. Hare and Nasci (1986) noted delayed mortality in surviving larvae of *Ae. aegypti* exposed to a median lethal concentration of Bti. Mulla and Singh

(1991) examined some biological parameters and morphogenetic aberrations of *Culex quinquefasciatus* Say larvae, pupae, and adults after treating larvae with sublethal concentrations of Bti.

Recent studies have demonstrated the potential of entomopathogenic fungi to control mosquito vectors (Farenhorst et al. 2008; Mnyone et al. 2009). These fungi do not cause instant mortality, but cause sublethal and later-life lethal effects on different stages of the mosquito life cycle. Due to such properties, fungi can be potentially used as “evolution-proof” agents and overcome mosquito resistance, unlike the currently deployed fast-acting chemical insecticides (Mnyone et al. 2011).

Leptolegnia chapmanii Seymour (Straminipila: Peronosporomycetes) has demonstrated marked pathogenicity toward the larvae of a number of mosquito species, with little or no effect on non-target insects (López Lastra et al. 2004). It is important to note that until 2005, *L. chapmanii* was considered an aquatic fungi belonging to Phylum Oomycota.

In the past few years, the strain LPSC 1099-ARSEF 5499 of *L. chapmanii* has shown encouraging results with regard to its pathogenicity on *Ae. aegypti*. In previous works, we studied biotic and abiotic factors affecting the infection with *L. chapmanii* (Pelizza et al. 2007a, b), and also deepened the understanding of longevity and infectivity of zoospores (Pelizza et al. 2008), the production of oogonia and oospores at different temperatures (Pelizza et al. 2010), and the combination of *L. chapmanii* zoospores with other larvicides such as Bti and temephos (Pelizza et al. 2010).

Therefore, the aim of this study was to determine the sublethal effects of *L. chapmanii* on fecundity, the number of gonotrophic cycles, fertility, and the relationship between wing length and fecundity in *Ae. aegypti* females.

Materials and Methods

Mosquito larvae

The *Ae. aegypti* larvae used in this study were obtained from colonies maintained following standard mosquito-rearing techniques (Gerberg et al. 1994).

Pathogen culture

The *L. chapmanii* strain used in this study (LPSC 1099-ARSEF 5499) was obtained from a puddle with infected larvae of *Ochlerotatus albifasciatus* (Macquart) in the city of Melchor Romero, La Plata, Buenos Aires province, Argentina (López Lastra et al. 1999). The *L. chapmanii* strain LPSC 1099-ARSEF 5499 was maintained on Emerson's YpSS agar medium (yeast extract 4 g, HK₂PO₄ 1 g, MgSO₄ 0.5 g, starch 15 g, agar 20 g, distilled water 1000 mL) in 60 x 15 mm sterilized Petri dishes. The zoospore inoculum was obtained as previously by Pelizza et al. (2011).

Number of gonotrophic cycles

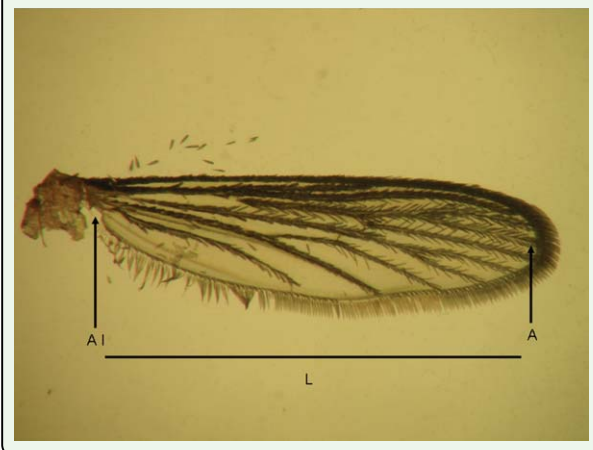
To study the possible sublethal effects of *L. chapmanii* on *Ae. aegypti* females that survived infection with zoospores of *L. chapmanii*, 200 healthy *Ae. aegypti* larvae in the third and fourth stages and a dose of 2.2×10^6 zoospores/mL of *L. chapmanii* were placed in five 1000-mL plastic containers with 800 mL of distilled water, as in a previous experiment by Pelizza et al. (2008). Additionally, 200 healthy larvae of *Ae. aegypti* in the third and fourth stages were placed in five plastic containers like to those described above and used as controls. Treated

and control containers were placed in incubators at 25° C and with a 12:12 L:D photoperiod. At 24 hr, the containers were examined and pupae were placed individually in glass tubes (3.5 cm in diameter x 7.5 cm in height) with a top covered with wire mesh. Then, 4 mL of distilled water was placed in each tube and a raisin was placed on the mesh as a source of carbohydrates for adults. Treated and control pupae were maintained at $27 \pm 1^\circ$ C, 80% relative humidity, and 12:12 L:D. A total of 200 treated insect (100 females and 100 males) and 200 control insect (100 females and 100 males) were placed in different cages covered with a wire net (50 x 50 x 50 cm) for 72 hr, which allowed the male and female mosquitoes to mate. After this time, an immobilized hen was placed in each cage for 1 hr as a source of blood for mosquito females. After 73 hr, fed females were placed individually in glass tubes with wet paper placed in the inner perimeter to allow the females to lay eggs on the moist paper.

Treated and control females were then kept at $27 \pm 1^\circ$ C, 50% relative humidity, and 12:12 L:D. After oviposition, between 72 and 96 hr after ingestion of blood, eggs were counted and treated and control females that survived were returned to their respective cages. This procedure was repeated until all *Ae. aegypti* females (both treated and controls) died, and these data were used to evaluate the number of gonotrophic cycles, defined as the period between feeding the female, egg laying, and the next feeding. The experiments described above were replicated three times on different dates under similar laboratory conditions.

The numbers of eggs laid by treated and control females in each gonotrophic cycle were compared by one-way ANOVA test (raw data after checking for homogeneity of variances and normality).

Figure 1. Wing of *Aedes aegypti* female. (AI) axillary incision; (A) apex; (L) maximum length. High quality figures are available online.



Relationship between fecundity and wing length

Forewings were measured in dead females (treated and control), following the method of Packer and Corbet (1989). Both wings were measured, and the larger measurement was used for the analysis. The maximum length was determined from the axillary incision to the apex of the wing, excluding the marginal fringe (Figure 1). The experiments described above were replicated three times on different dates under similar laboratory conditions.

The wing length of treated and control females were compared by one-way ANOVA test (raw data after checking for homogeneity of variances and normality).

Fertility

To investigate possible sublethal effects on fertility (number of viable eggs), moist papers with eggs laid in each gonotrophic cycle by both treated and control females of *Ae. aegypti* were immersed in different plastic trays (38 x 28 cm) with 4000 mL of dechlorinated water and finely ground fish as food. Trays containing the eggs laid by treated and control females were maintained at $27 \pm 1^\circ \text{C}$ and 12:12 L:D. First-stage larvae were counted

counted and removed daily for five days. The experiments described above were replicated three times on different dates under similar laboratory conditions.

The number of viable eggs laid by treated and control females was analyzed by one-way ANOVA test and transformed by $\log(x+1)$ function to achieve homogeneity and normality.

Results and Discussion

Significant differences were observed in the fecundity of females of *Ae. aegypti* exposed to zoospores of *L. chapmanii* compared to the controls (Table 1). Significant differences were also observed in the number of gonotrophic cycles of females of *Ae. aegypti* that survived *L. chapmanii* infection as compared to the controls (six cycles vs. eight cycles; Table 1). Significant differences (ANOVA, $F = 110.90$, $df = 599$, $p < 0.0001$) were also observed in the maximum length of the wing ($1.96 \text{ mm} \pm 0.59$ in *Ae. aegypti* females that had been in contact with *L. chapmanii* zoospores vs. $2.32 \pm 0.18 \text{ mm}$ in controls).

Significant differences were observed in fertility between females of *Ae. aegypti* exposed to zoospores of *L. chapmanii* and the controls (Table 2). The total percentage of viable eggs laid by treated *Ae. aegypti* females in their six gonotrophic cycles was $41.76 \pm 5.95 \%$, whereas the percentage of viable eggs laid by control females was $70.23 \pm 6.79 \%$ in their

Table 1. Results of the ANOVA and the average number of eggs laid by treated and control *Aedes aegypti* females for each gonotrophic cycle.

Gonotrophic cycles	Treated		Control		DF	F-value	p
	Mean \pm SD	n	Mean \pm SD	n			
1	61.3 \pm 10	300	84.10 \pm 11.83	300	599	210.38	1E-04
2	59.5 \pm 7.37	238	80.67 \pm 9.14	268	505	164.11	1E-04
3	56.23 \pm 8.54	190	75.55 \pm 4.78	232	421	95.48	1E-04
4	53.64 \pm 6.61	151	72.49 \pm 10.69	186	336	74.94	1E-04
5	48.2 \pm 12.17	92	70.85 \pm 8.11	147	238	81.08	1E-04
6	47.26 \pm 4.62	61	69.93 \pm 13.2	111	171	47.54	1E-04
7	-	-	68.32 \pm 11.78	73	-	-	-
8	-	-	66.35 \pm 9.56	48	-	-	-

*73 and 48 *Ae. aegypti* control females reached the seventh and the eighth gonotrophic cycle respectively.

Table 2. Results of the ANOVA and the percentage of hatched eggs laid by treated and control *Aedes aegypti* females in each gonotrophic cycle.

Gonotrophic cycles	Treated Females			Control Female			DF	F-value	p
	Eggs laid	Hatched eggs	Percentage of hatched eggs \pm SD	Eggs laid	Hatched eggs	Percentage of hatched eggs \pm SD			
1	18,554	6,423	34.62 \pm 6.42	25,229	14,081	55.66 \pm 10	5	4.44	0.103
2	14,168	6,373	44.98 \pm 7	21,622	16,504	76.33 \pm 11.06	5	14.43	0.019
3	10,696	4,599	43 \pm 9.16	17,522	14,466	82.56 \pm 2.51	5	56.22	0.002
4	8,090	2,939	36.33 \pm 6.49	13,482	9,437	70 \pm 4	5	55.27	0.002
5	4,441	2,263	50.97 \pm 3.6	10,421	8,719	83.67 \pm 5.12	5	65.75	0.001
6	2,884	1,173	40.66 \pm 3.05	7,762	5,640	72.66 \pm 2.51	5	68.89	0.001
7	-	-	-	4,498	2,578	57.31 \pm 7.03	-	-	-
8	-	-	-	3,186	2,028	63.65 \pm 6.11	-	-	-

eight gonotrophic cycles.

At the time of this study, the known world distribution of *L. chapmanii* is restricted to three states of the USA (California, Florida, and Ohio) and the city of Melchor Romero, La Plata, Buenos Aires province, Argentina (Mc Innis and Zattau 1982; Seymour 1984; Lord and Fukuda 1988; Fukuda et al. 1997; López Lastra et al. 1999).

This study is the first to examine the sublethal effects of *L. chapmanii* on females of *Ae. aegypti* that survived infection with *L. chapmanii*. Females that survived infection with *L. chapmanii* zoospores had smaller forewings in length and laid fewer eggs than the controls. The results are coincident with those observed by Packer and Corbet (1989) in *Aedes punctator* (Kirby) and Blakmore et al. (2000) in *Aedes albopictus* (Skuse). This study observed the correlation between body size, wing size, and the number of eggs laid, and found that females with smaller wings and smaller body size had lower fertility.

Similar results were observed by Flores et al. (2004), who applied a sublethal concentration of Bti on *Ae. aegypti* and found a significant reduction in fecundity in females of this mosquito. Also, it has been shown that three pyrethroids (d-phenothrin, d-allevethrin, and tetramethrin) reduce the fecundity of *Ae. aegypti* when applied in sublethal doses (Shaalan et al.

2005). Scholte et al. (2006) observed a fecundity reduction in samples of *Anopheles gambiae* Giles treated with the entomopathogenic fungus *Metarhizium anisopliae*.

It is important to note that we observed reduced survival and therefore a reduced number of gonotrophic cycles in *Ae. aegypti* females exposed to zoospores of *L. chapmanii*. Similar results were obtained by Flores et al. (2004), who treated *Ae. aegypti* with a sublethal dose (CL70) of Bti. Mnyone et al. (2011) observed a significant reduction in survival of *An. gambiae* treated with the entomopathogenic fungi *Beauveria bassiana* and *M. anisopliae*. We observed a significant reduction in fertility (number of viable eggs) of females of *Ae. aegypti* that survived infection with zoospores of *L. chapmanii* when compared with control females. Using different sublethal doses of Bti (CL30, CL50, and CL70), Flores et al. (2004) obtained similar results, i.e., a greater reduction in survival, fecundity, and fertility in *Ae. aegypti* females when increasing the dose of this biocontrol agent. Belinato et al. (2009) evaluated *Ae. aegypti* larvae with a sublethal dose of triflumuron, a chitin synthesis inhibitor, and observed a reduction in fertility in surviving mosquito females when compared with control females.

This study marks the first time that the effects of *L. chapmanii* have been determined on *Ae.*

aegypti larvae surviving *L. chapmanii* zoospore infection. This information is relevant because the decrease in survival of treated females leads to a smaller number of gonotrophic cycles. Reduced fecundity, body size (smaller length of forewings), and fertility (number of viable eggs) were also observed in treated females. Therefore, *L. chapmanii* not only causes a high and quick mortality in *Ae. aegypti* larvae, as shown in previous studies (Pelizza et al. 2008), but also reduces the reproductive capacity of larvae that survive the infection.

Although it is important to perform field studies to evaluate *L. chapmanii* as a control agent more fully, the characteristics of this control agent (high mortality and reduced reproductive capacity) seem to create a significant reduction in the population of *Ae. aegypti* in the mid and long term, and consequently reduce the health risks caused by *Ae. aegypti* as a vector of diseases.

Acknowledgments

This study was partially supported by CICPBA, CONICET, and Universidad Nacional de La Plata, La Plata, Argentina.

References

Adugelo-Silva F, Spielman A. 1984. Paradoxical effects of simulated larviciding on production of adult mosquitoes. *American Journal of Tropical Medicine and Hygiene* 88: 1267–1269.

Belinato TA, Martins AJ, Lima JBP, Camara TN, Peixoto AA, Valle D. 2009. Effect of the chitin synthesis inhibitor triflumuron on the development, viability and reproduction of *Aedes aegypti*. *Memórias do Instituto Oswaldo Cruz* 104: 43–47.

Blackmore MS, Lord CC. 2000. The relationship between size and fecundity in *Aedes albopictus*. *Journal of Vector Ecology* 25: 212–217.

Farenhorst M, Farina D, Scholte EJ, Takken W, Hunt RH, Coetzee M, Knols BGJ. 2008. African water storage pots for the delivery of the entomopathogenic fungus *Metarhizium anisopliae* to the malaria vectors *Anopheles gambiae* s.s. and *Anopheles funestus*. *American Journal of Tropical Medicine and Hygiene* 78: 910–916.

Flores AE, Garcia GP, Badil MH, Tovar LR, Salas IF. 2004. Effects of sublethal concentrations of vectovac® on biological parameters of *Aedes aegypti*. *Journal of American Mosquito Control Association* 20: 412–417.

Fukuda T, Willis OR, Barnard DR. 1997. Parasites of the Asian Tiger mosquito and other container-inhabiting mosquitoes (Diptera: Culicidae) in northcentral Florida. *Journal of Medical Entomology* 84: 226–233.

Gerberg E, Barnard D, Ward R. 1994. Manual for mosquito rearing and experimental techniques. *American Mosquito Control Association Bulletin* 5: 1–98.

Hare SGF, Nasci RS. 1986. Effects of sublethal exposure to Bti on larval development and adult size in *Aedes aegypti*. *Journal of American Mosquito Control Association* 2: 325–328.

Koella JC, Lynch PA, Thomas MB, Read AF. 2009. Towards evolution-proof malaria control with insecticides. *Evolutionary Applications* 2: 469–480.

- López Lastra CC, Steciow MM, García JJ. 1999. Registro más austral del hongo *Leptolegnia chapmanii* (Oomycetes: Saprolegniales) como patógeno de larvas de mosquitos (Diptera: Culicidae). *Revista Iberoamericana de Micología* 16:143–145.
- López Lastra CC, Scorsetti AC, Marti GA, García JJ. 2004. Host range and specificity of an Argentinean isolate of the aquatic fungus *Leptolegnia chapmanii* (Oomycetes: Saprolegniales), a pathogen of mosquito larvae (Diptera: Culicidae). *Mycopathologia* 158: 311–315.
- Lord JC, Fukuda T. 1988. An ultrastructural study of *Culex quinquefasciatus* larvae by *Leptolegnia chapmanii* (Oomycetes: Saprolegniales). *Mycopathologia* 104: 67–74.
- Mnyone LL, Kirby MJ, Lwetoijera DW, Mpingwa MW, Knols BGJ, Takken W, Russell TL. 2009. Infection of the malaria mosquito, *Anopheles gambiae* with two species of entomopathogenic fungi: effects of concentration, co-formulation, exposure time and persistence. *Malaria Journal* 8: 309.
- Mnyone LL, Kirby MJ, Mpingwa MW, Lwetoijera DW, Knols BGJ, Takken W, Koenraadt CJM, Russell TL. 2011. Infection of *Anopheles gambiae* mosquitoes with entomopathogenic fungi: effect of host age and blood-feeding status. *Parasitology Research* 108: 317–322.
- Mc Innis T, Zattau WC. 1982. Experimental infection of mosquito larvae by a species of the aquatic fungus *Leptolegnia*. *Journal of Invertebrate Pathology* 39: 98–104.
- Mulla MS, Singh N. 1991. Delayed mortality and morphogenetic anomalies induced by the microbial control agent *Bacillus thuringiensis* ser (H14) in *Culex quinquefasciatus*. *Journal of American Mosquito Control Association* 7: 420–423.
- Ochoa G, Arrivillaga J. 2009. *Bacillus thuringiensis*: Avances y perspectivas en el control biológico de *Aedes aegypti*. *Boletín de malariología y salud ambiental* 59: 181–191.
- Packer MJ, Corbet PS. 1989. Size variation and reproductive success of female *Aedes punctator* (Diptera: Culicidae). *Ecological Entomology* 14: 297–309.
- Pelizza SA, López Lastra CC, Becnel JJ, Bisaro V, García JJ. 2007a. Biotic and abiotic factors affecting *Leptolegnia chapmanii* infection in *Aedes aegypti* L. (Diptera: Culicidae). *Journal of American Mosquito Control Association* 23: 177–181.
- Pelizza SA, López Lastra CC, Becnel JJ, Bisaro V, García JJ. 2007b. Effects of temperature, pH and salinity on the infection of *Leptolegnia chapmanii* Seymour (Peronosporomycetes) in mosquito larvae. *Journal of Invertebrate Pathology* 96: 133–137.
- Pelizza SA, López Lastra CC, Becnel JJ, Humber RA, García JJ. 2008. Further research on the production, longevity and infectivity of the zoospores of *Leptolegnia chapmanii* Seymour (Oomycota: Peronosporomycetes). *Journal of Invertebrate Pathology* 98: 314–319.
- Pelizza SA, Scorsetti AC, López Lastra CC, García JJ. 2010. Production of oogonia and oospores of *Leptolegnia chapmanii* Seymour (Straminipila: Peronosporomycetes) in *Aedes aegypti* (L.) larvae at different temperatures. *Mycopathologia* 169: 71–74.

Pelizza SA, Scorsetti AC, Bisaro V, López Lastra CC, García JJ. 2010. Individual and combined effects of *Bacillus thuringiensis* var. *israelensi*, *Temephos* and *Leptolegnia chapmanii* on the larval mortality of *Aedes aegypti*. *Biocontrol* 55: 647–656.

Pelizza SA, Cabello MN, Tranchida MC, Scorsetti AC, Bisaro V. 2011. Screening for a culture medium yielding optimal colony growth, zoospore yield and infectivity of different isolates of *Leptolegnia chapmanii* (Straminipila: Peronosporomycetes). *Annals of Microbiology* 61: 991–997.

Scholte EJ, Knols BG, Takken W. 2006. Infection of the malaria mosquito *Anopheles gambiae* with the entomopathogenic fungus *Metarhizium anisopliae* reduces blood feeding and fecundity. *Journal of Invertebrate Pathology* 91: 43–49.

Seymour RL. 1984. *Leptolegnia chapmanii*, an Oomycete pathogen of mosquito larvae. *Mycologia* 76: 113–120.

Shaalan EAS, Canyon DV, Younes MWF, Wahab HA, Mansour AH. 2005. Effects of sublethal concentrations of synthetic insecticides and *Callitris glaucophylla* extracts on the development of *Aedes aegypti*. *Journal of Vector Ecology* 30: 295–298.

Simsek FM, Akiner MM, Caglar SS. 2009. Effects of sublethal concentration of Vectobac 12 AS on some biological parameters of the malaria vector *Anopheles superpictus*. *Journal of Animal and Veterinary Advances* 8: 1326–1331.

Zaim M, Guillet P. 2002. Alternative insecticides: an urgent need. *Trends in Parasitology* 18: 161–163.