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Source: Zoological Science, 15(6) : 861-869

Published By: Zoological Society of Japan

URL: <https://doi.org/10.2108/zsj.15.861>

# Identification of Two Distinct Muscles in the Planarian *Dugesia japonica* by their Expression of Myosin Heavy Chain Genes

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**ABSTRACT**—Ultrastructural and physiological studies have shown that planarian muscles have some characteristics of smooth and some characteristics of striated muscles. To characterize planarian muscles, we isolated two myosin heavy chain genes (*DjMHC-A* and *DjMHC-B*) from a planarian, *Dugesia japonica*, by immunological screening, and analyzed their structures and spatial expression patterns. Structural analysis indicated that both MHC genes are striated-muscle-type myosin genes, although planarian muscles do not have any striation. *In situ* RNA hybridization showed that expression of the two myosin genes is spatially strictly segregated. *DjMHC-A* was expressed in pharynx muscles, pharynx cavity muscles, muscles surrounding the intestinal ducts, a subpopulation of body-wall muscles and several muscle cells in the mesenchymal region around the base of the pharynx. *DjMHC-B* was expressed in body-wall muscles (including circular, diagonal and longitudinal muscles), vertical muscles and horizontally oriented muscles. Double staining with *DjMHC-A* and *-B* probes clearly demonstrated that expression of the *DjMHC-A* and *-B* genes do not occur in the same cell. During regeneration, the number of cells positive for expression of each gene increased in the blastema region, suggesting that both types of muscle may be involved in blastema formation. *DjMHC-B*-positive cells disappeared from the body-wall muscle layer in the pharynx-cavity-forming region, whereas *DjMHC-A*-positive cells were markedly accumulated there, suggesting that the two types of muscle in the body wall layer may have distinct functions. These results indicate that planarians have at least two types of muscle that express striated-muscle-type MHC genes, but do not form striation.

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## INTRODUCTION

The muscles of most invertebrates and vertebrates can be clearly classified as striated or smooth muscles by their morphological characteristics. However, oblique muscles, which occur in some invertebrates, have characteristics intermediate between those of striated and smooth muscles (Hanson, 1956; Lowy and Hanson, 1962), i.e., they contain dense bodies characteristic of smooth muscles although they have helical striation composed of myofilaments. Muscles of planarians have even more ambiguous characteristics (MacRae, 1963; Morita, 1965; Sarnat, 1984). Their structure resembles that of smooth muscle, since they have no striation and contain dense bodies, but their nuclei are located in peripheral regions of the cells, which is characteristic of striated muscles, and a Z-line-like structure directly connected to sarcolemma is also observed (Hori, 1983). We have therefore begun to clarify the nature of planarian muscles by using cloned genes to probe the expression of muscle-specific

genes.

It is well known that many organisms have multiple isoforms of myosin heavy chains (MHCs), which are classified into three types by their structure and expression pattern: striated muscle, smooth muscle and nonmuscle types (Warrick and Spudich, 1987). Thus, it is expected that MHC genes should be good landmark genes to classify morphologically ambiguous planarian muscles at the molecular level. A short fragment of an MHC gene of a planarian, *Girardra tigrina*, has already been cloned by immunological screening using a muscle-specific monoclonal antibody (Bueno *et al.*, 1997; Cebria *et al.*, 1997). However, only one MHC gene has been cloned so far using that method, and the cloned cDNA fragment is too short to fully characterize the MHC gene(s) of planarians. Here we report the structure and expression patterns of two different MHC genes isolated from the planarian *Dugesia japonica*, and describe some resultant insights into the characteristics and evolution of planarian muscles.

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## MATERIALS AND METHODS

### Animals

A clonal strain of planarian, *Dugesia japonica*, established in our laboratory, was used. It was bred in boiled tap water at 22–24°C, and fed chicken liver twice a week. All worms were used for experiments after one week of starvation.

### Cloning and DNA sequencing

Mouse antiserum against living planarians was used to screen a  $\lambda$ ZAP expression cDNA library of planarian. Immunological screening was performed according to the manufacture protocols (Stratagene). Forty-seven positive clones among  $4 \times 10^4$  cDNA clones were classified by cross-reactivity and restriction enzyme analyses. These positive clones were recloned into pBluescript by *in vivo* excision. Purified plasmid DNA was sequenced using a BcaBEST Dideoxy Sequencing Kit (Takara) or a Thermo Sequence Fluorescent Labeled Primer Cycle Sequencing Kit (Amersham). Analysis of the sequence was performed using the DDBJ on-line service (National Institute of Genetics, Japan) or the Wisconsin GCG Package program (Version 9.0, Genetics Computer Group, Madison, WI, USA).

### In situ hybridization of sections and whole-mount specimens

Planarians were treated with 2% HCl for 5 min and fixed either in relaxant solution (1% HNO<sub>3</sub>, 1.6% formaldehyde, 20  $\mu$ M MgSO<sub>4</sub>; Dawar, 1973) for 12–24 hr, or in 2% formaldehyde, 0.2% saturated picric acid solution for 30 min, followed by 2% formaldehyde, 0.2% saturated picric acid solution, 0.1% glutaraldehyde for 1 hr, and then 2% formaldehyde, 0.2% saturated picric acid solution for 2 hr. They were then dehydrated through a graded ethanol series, embedded in paraffin, sectioned horizontally, sagittally or transversely and then mounted on poly-L-lysine-coated slides. After de-paraffination in xylene and hydration through the ethanol series, the mounted specimens were rinsed in PBS and acetylated in 0.1 M triethanolamine, 0.09% HCl and 0.25% acetic anhydride for 15 min. They were rinsed in  $2 \times$  SSC and then incubated at 55°C for 30 min in 50% formamide,  $5 \times$  SSC and 10 mM DTT. Digoxigenin-labeled antisense probe derived from *DjMHC-A* or *DjMHC-B* was diluted to 50 ng–500 ng/ml in 50% formamide,  $5 \times$  SSC, 0.1 mg/ml yeast RNA, 0.1 mg/ml heparin, 0.1% Tween-20, 10 mM DTT and 10% dextran sulfate, denatured at 65°C for 15 min, and added to slides for hybridization at 55°C for 16–36 hr. After hybridization, the slides were washed in 50% formamide,  $5 \times$  SSC and 0.1% Tween-20 for 9–12 hr at 55°C, and twice in  $2 \times$  SSC at room temperature for 15 min. The slides were rinsed in Buffer-1 (0.1 M maleic acid, 0.15 M NaCl, 0.1% TritonX-100), incubated in Buffer-2 (1% Boehringer blocking reagent in Buffer-1) for 30 min and then in Buffer-2 containing alkaline-phosphatase (AP)-conjugated anti-digoxigenin antibody (Boehringer) diluted 1:5000 at room temperature for 2 hr. They were rinsed in Buffer-1 and in TMN buffer (0.1 M NaCl, 0.1 M Tris-HCl, pH 9.5, 0.05 M MgCl<sub>2</sub>) for 15 min. Signals were rendered detectable by incubation of the slides in TMN buffer containing 200  $\mu$ g/ml NBT (Sigma) and 175  $\mu$ g/ml BCIP (Sigma) for 2–3 hr. After the chromogenic reaction was completed, the slides were washed twice in TE buffer (10 mM Tris-HCl, pH 8, 1 mM EDTA), mounted in 50% glycerol in TE buffer and observed under a microscope. Double staining was performed using digoxigenin-labeled probe for MHC-B and biotin-labeled probe for MHC-A. Both signals were simultaneously detected using a TSA Kit (NEN Life Science Products) according to the procedures of Kato *et al.* (manuscript in preparation). Whole-mount *in situ* hybridization was carried out as described by Umesonon *et al.* (1997) and Agata *et al.* (1998).

### Dot blot hybridization

Non-labeled sense RNAs synthesized from *DjMHC-A* or *DjMHC-B* and yeast tRNA were diluted serially and blotted on nylon membranes (Hybond-N, Amersham) in duplicate. The RNA was fixed to the membrane by baking at 80°C for 2 hr and cross-linking by UV

light. Both membranes were incubated in prehybridization solution at 55°C for 1 hr. They were then incubated with the probe at 50 ng/ml at 55°C for 16–36 hr. After hybridization, they were washed in the wash buffer at 55°C for 9–12 hr and washed twice in  $2 \times$  SSC at room temperature for 15 min. The hybridization signal was detected immunologically as described above.

## RESULTS

### Isolation and characterization of two planarian MHC genes

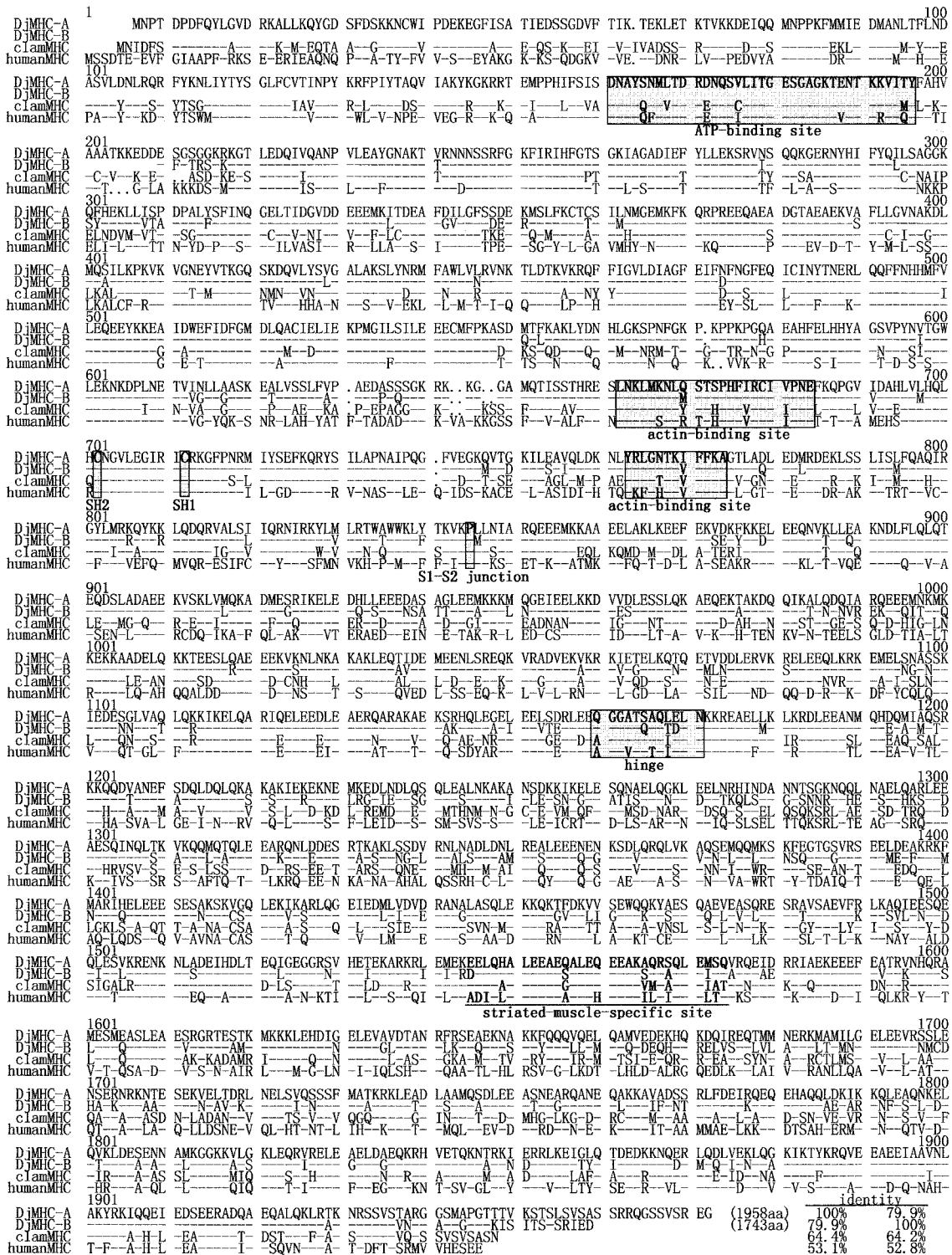
Forty-seven clones that reacted with anti-planarian antiserum were classified into two groups (A and B) by cross-reactivity and restriction enzyme analyses. The longest clone of group A contained a 6111-bp insert with an ORF of 1958 amino acids. The longest clone of group B contained a 5388-bp insert with an ORF of 1743 amino acids. The nucleotide sequence data reported in this paper will appear in the DDBJ/EMBL/GenBank nucleotide sequence databases with the accession numbers AB015484 and AB015485.

Computer analysis of the DNA and amino acid sequences showed high similarity between the two genes (79.9% at the protein level) and similarity of both genes to myosin heavy chain genes of other animals (Fig. 1). The predicted amino acid sequence of the group-A gene shows 64.4% similarity to that of scallop and 53.1% similarity to that of human MHC. That of the group-B gene shows 64.2% and 52.8% similarity to those of scallop and human MHCs, respectively. Myosin-specific domains such as the ATP-binding site, actin-binding sites, two closely spaced reactive thiol groups; SH1 and SH2, and hinge were well conserved. Therefore, we named the group-A and -B genes "*DjMHC-A*" and "*DjMHC-B*", respectively.

To classify these two planarian MHCs, we compared them to striated-muscle-type and smooth-muscle-type MHCs of other organisms. Figure 2A shows a comparison of the amino acid sequences of the isoform-specific region among *DjMHC-A*, *DjMHC-B* and MHCs of various other organisms (Schuchert *et al.*, 1993; Takahashi *et al.*, 1992). This comparison indicated that both *DjMHC-A* and *DjMHC-B* genes are striated-muscle-type MHC genes since they showed much higher similarities to striated-muscle-type than to smooth- or nonmuscle-type MHCs. Phylogenetic tree analysis also suggested that both planarian MHC genes should be classified as striated-muscle type. *DjMHC-A* and *DjMHC-B* are related more closely to each other than to any other MHC gene. Although these structural analyses suggest that both MHCs are striated-muscle-type MHCs, no striation has been observed in planarian muscles (MacRae, 1963; Morita, 1965; Hori, 1983).

### Comparison between *DjMHC-A* and *DjMHC-B*

We next compared the structures of *DjMHC-A* and *DjMHC-B*. The overall identities between *DjMHC-A* and *DjMHC-B* are 74.9% and 79.9% at the DNA and protein levels, respectively. HarrProt analysis clearly indicated regions conserved between *DjMHC-A* and *DjMHC-B* (Fig. 3A). The head region is highly homologous between them (92.2%), but the light meromyosin (LMM) region of the tail is much less

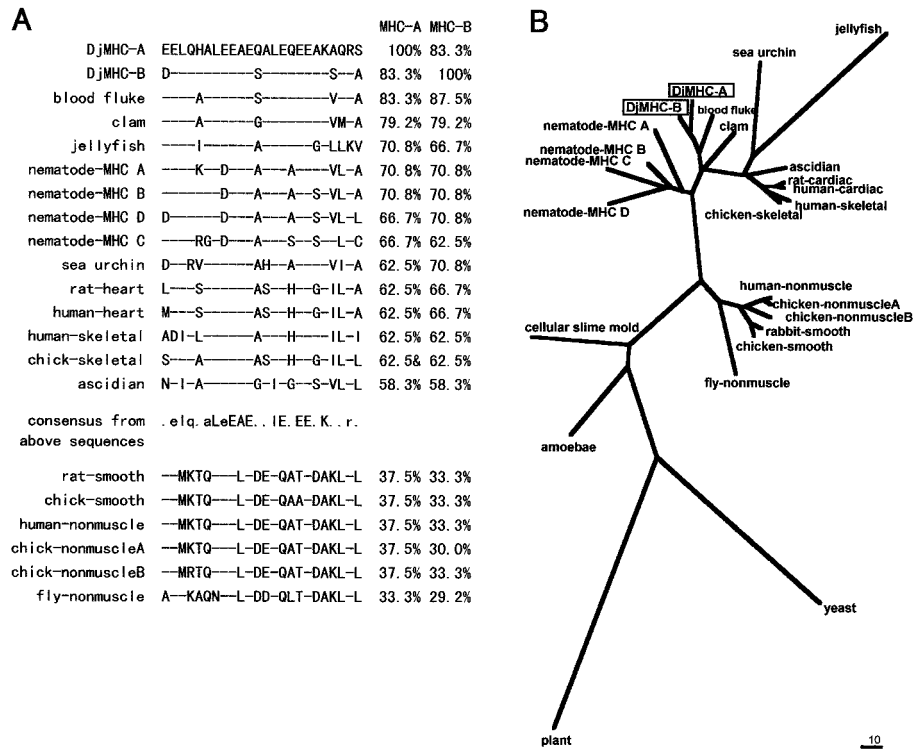


**Fig. 1.** Predicted amino acid sequences of *DjMHC-A* and *DjMHC-B* compared to those of clam (scallop) and human MHCs. Conserved amino acids are indicated by dashes. Dots indicate gaps for alignment. Myosin-specific domains of the MHCs are boxed. Underlines indicate amino acid sequences conserved among the striated-muscle MHCs.

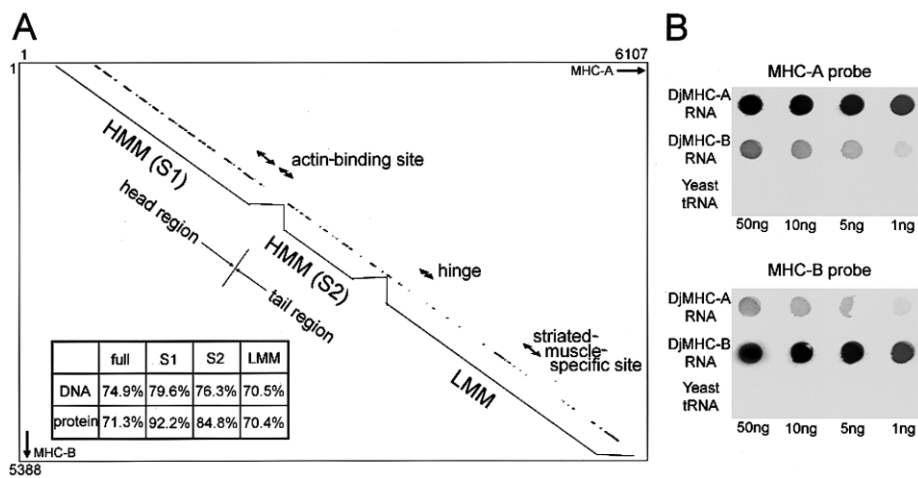
homologous (70.4%). Since the two genes show strong homology in the head region, we detected cross-reaction between them during routine hybridization procedures. Thus, we needed to establish conditions for avoiding cross-reaction

during *in situ* hybridization. First, we used tail regions as probes, since these regions have low homology. However, the resultant hybridization signals were weaker than the signals obtained using full-length probes (data not shown). There-





**Fig. 2.** Comparison of amino acid sequences in the isoform-specific region, and phylogenetic tree analysis. (A) Alignment of the sequences in the isoform-specific region (Schuchert *et al.*, 1993). Complete sequences were aligned by the pileup program of the GCG Package and then the isoform-specific region was selected. Sequences were taken from the SWISS Plot and DDBJ/GenBank/EMBL sequence libraries. Dashes indicate identical residues. The consensus sequence of the striated-muscle-type MHCs is indicated. Capital letters indicate perfect conservation; lower case letters, high conservation; points, no conservation. (B) An unrooted phylogenetic tree of DjMHC-A, DjMHC-B and other MHCs. This tree was produced using the distances program of the GCG Package. The accession numbers and amino acid residues of the MHCs used in these analyses are as follows: amoebae, P05659; ascidian, D45163; blood fluke, L01634; cellular slime mold, P08799; chicken-skeletal, P13538; chicken-nonmuscleA, P14105; chicken-nonmuscleB, M93676, PID g212449; chicken-smooth, P10587; clam, P24733; fly-nonmuscle, Q99323; human-nonmuscle, P35579; human-skeletal, P11055; human-cardiac, P12883; jellyfish, X64057; nematode-MHC A, P12844; nematode-MHC B, P02566; nematode-MHC C, P12845; nematode-MHC D, P02567; plant, Z34293; rabbit-smooth, P35748; rat-cardiac, P02563; sea urchin, M55321; yeast, P08964. Accession numbers that begin with a “P” are from the SWISS Plot database; all others are from the DDBJ/GenBank/EMBL database. “10” indicates the phylogenetic distance expressed as substitutions per 100 amino acids.



**Fig. 3.** Comparison between structures of *DjMHC-A* and *DjMHC-B*. (A) HarrProt analysis was performed to compare the *DjMHC-A* and *DjMHC-B* genes. Unit size used for the comparison: 30. Dot plot matching number: 25. HMM, heavy meromyosin; LMM, light meromyosin; S1, subunit 1; S2, subunit 2. The table summarizes the identity between *DjMHC-A* and *DjMHC-B* in three regions. (B) Cross-reactivity between *DjMHC-A* and *DjMHC-B* in high stringency condition. The sense RNAs of *DjMHC-A* and *DjMHC-B*, and yeast tRNA were diluted serially and blotted on nylon membranes. Dot blot hybridization was performed using labeled antisense RNA probes: *DjMHC-A* (upper panel), *DjMHC-B* (lower panel).

fore, we tried performing RNA hybridization at 55°C in a solution containing 50% formamide, and checked the cross-reactivity between *DjMHC-A* and *DjMHC-B* by serial dilution experiments (Fig. 3B). We obtained satisfactory results using these high stringency conditions, although slight cross-reactivity was detected.

### Expression of *DjMHC-A* and *DjMHC-B* in normal adult planarians

To investigate the spatial distribution of planarian MHC gene transcripts in normal adult planarians, we performed *in situ* hybridization under the high stringency conditions described above. Strong expression of *DjMHC-A* was detected in pharynx muscles (Fig. 4A-a, b and d). *DjMHC-A*-positive signals were also observed in muscles surrounding the intestinal ducts (Fig. 4A-c and d; arrows), a subpopulation of body-wall muscles (Fig. 4A-c; arrowheads), several cells in the mesenchymal region around the base of the pharynx (Fig. 4A-c; asterisks), and pharynx-cavity muscles (Fig. 4A-d; arrowheads).

In contrast to the pattern seen with *DjMHC-A*, expression of *DjMHC-B* was detected in body-wall muscles (Fig. 4B-a, b and c; arrows), vertical muscles (Fig. 4B-a and b; arrowheads) and horizontally oriented muscles in the ventral part of the body (Fig. 4B-c; short arrows), but not in pharynx muscles. Interestingly, we found that the nucleus of the vertical muscles is located in the dorsal region (Fig. 4B-c; open arrowheads), whereas the nucleus of the horizontally oriented muscles is located in the ventral region (Fig. 4B-c; arrowheads).

Since we detected expression of both genes in the body-wall-muscle layer and the mesenchymal region, we performed doubly probed *in situ* hybridizations to investigate whether or not *DjMHC-A* and *DjMHC-B* were expressed in the same cells. In Fig. 5, *DjMHC-A*- and *DjMHC-B*-positive cells are shown in green and red colors, respectively. Blue color indicates nuclear counter-staining using Hoechst 33342. No overlapping expression was observed. In the mesenchymal region, *DjMHC-A*-positive cells were distributed at the periphery of the intestinal duct, but *DjMHC-B*-positive cells were observed in the space between the intestinal ducts (Fig. 5B-D). Figure 5E-G shows higher magnification views of the body-wall layer. *DjMHC-A* and *DjMHC-B* expression was observed in cells in the same layer, but not in the same cells. These results clearly demonstrate that muscle cells of planarians can be classified into at least two types by their patterns of expression of MHC genes.

### Expression of *DjMHC-A* and *DjMHC-B* in regenerating planarians

We analyzed the expression patterns of the two MHC genes during regeneration. Intact planarians were cut behind the pharynx, and the resultant regenerating planarians were analyzed by *in situ* hybridization. During regeneration, the cells expressing each MHC gene in the body-wall-muscle layer increased specifically in the blastema region (Fig. 6A and B; the blastema region is surrounded by dots). Furthermore, *DjMHC-B* expressing cells were also increased in the region

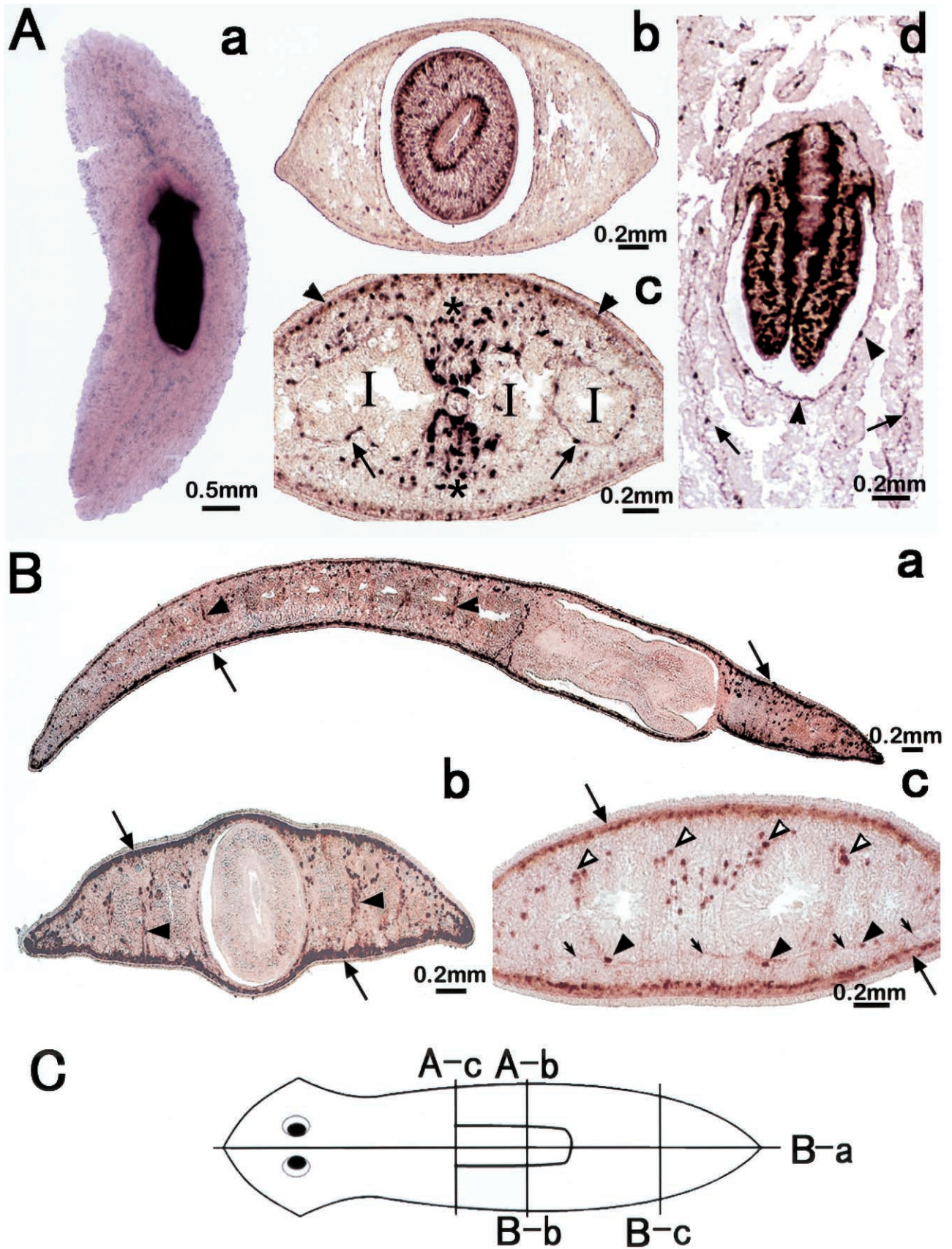
inside of the blastema region. Interesting views were obtained during the process of pharynx-cavity formation. After 5-6 days of regeneration, the pharynx cavity becomes open. Around that time, *DjMHC-B*-positive cells disappeared from the body-wall-muscle layer in the nascent cavity-forming region (Fig. 6D, asterisk), whereas *DjMHC-A*-positive cells were accumulated there (Fig. 6C, asterisk). Such complementary expression patterns suggest that *DjMHC-A*- and *DjMHC-B*-expressing cells may have distinct functions in the body-wall-muscle layer.

## DISCUSSION

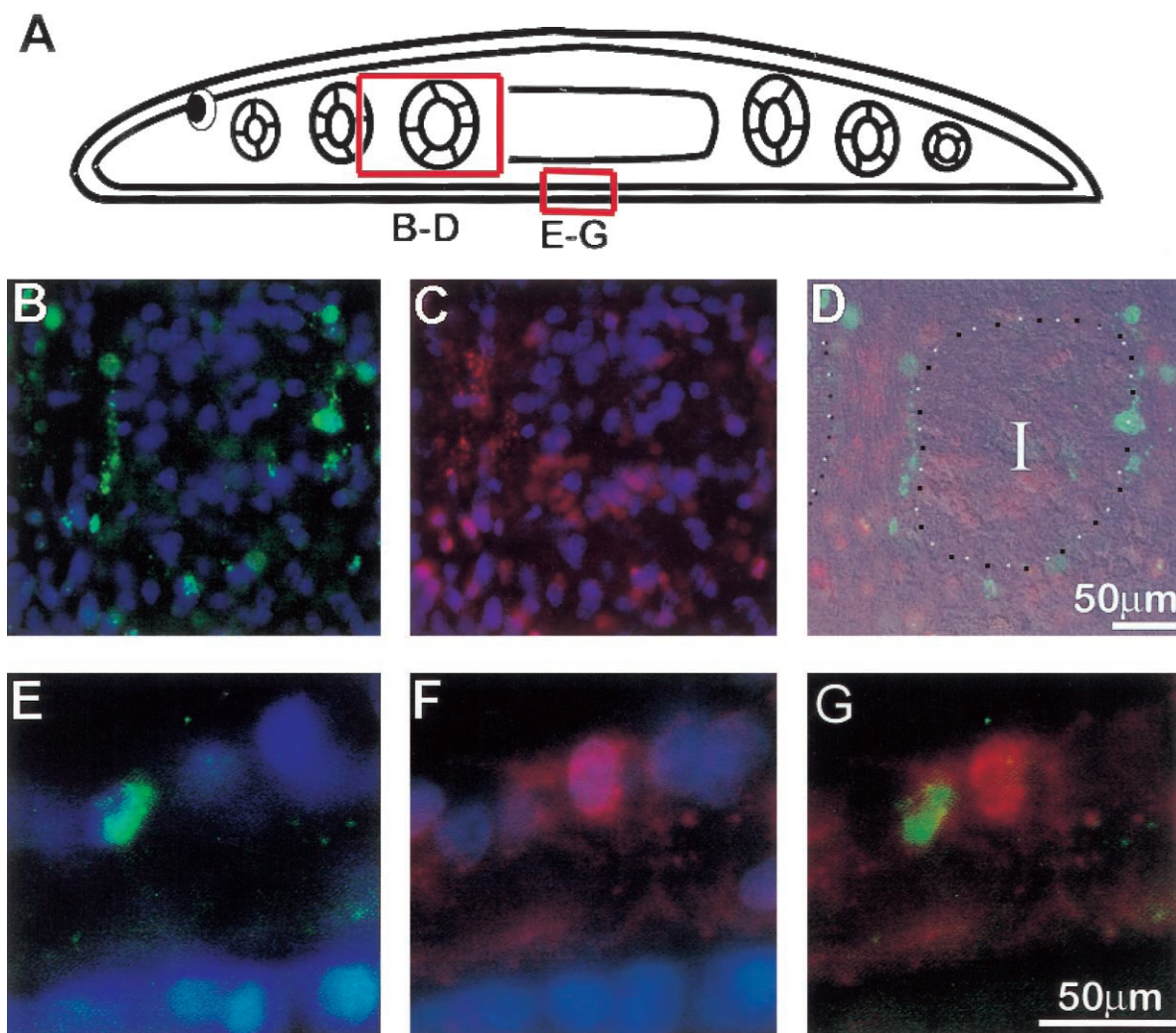
We have isolated two MHC genes, *DjMHC-A* and *DjMHC-B*, from a planarian, *Dugesia japonica*, and analyzed their structures and expression patterns. Surprisingly, both the *DjMHC-A* and *DjMHC-B* genes were of the striated-muscle type, although no striation was observed in planarian muscles (MacRae, 1963; Morita, 1965; Hori, 1983). We expected that the muscles surrounding the intestinal ducts might express an MHC gene of the smooth-muscle type, as in vertebrates. However, we were not able to isolate smooth-muscle-type MHC genes from planarian. The muscles surrounding the intestinal ducts express a striated-muscle-type MHC gene, *DjMHC-A*. In the case of *C. elegans*, four MHC genes have been isolated and all of them are classified as striated-muscle type: the MHC-A and -B genes are expressed in body-wall muscles, while the MHC-C and -D genes are expressed in pharynx muscles (Dibb *et al.*, 1988; Epstein *et al.*, 1974; Garcea *et al.*, 1978; Miller *et al.*, 1986; Fig. 2B). Morphological studies indicate that all the muscles of *C. elegans* are oblique muscles (Burr and Gans, 1998). These results obtained in planarians and nematodes suggest that all the muscle myosins of invertebrates may be encoded by striated-muscle-type MHC genes. Smooth muscles in vertebrates may have evolved from cells in other lineages, based on nonmuscle-type MHC genes.

Analysis of expression of *DjMHC-A* and *DjMHC-B* clearly indicated that planarian muscles are composed of at least two distinct types of muscles. *DjMHC-A* is specifically expressed in pharynx muscles, muscles surrounding the intestinal ducts, pharynx-cavity muscles and a subpopulation of body-wall muscles (Fig. 7A and B; green). On the other hand, *DjMHC-B* is expressed in body-wall muscles, vertical muscles and horizontally oriented muscles (Fig. 7A and B; red). These expression patterns suggest that the *DjMHC-A*-expressing muscles may have a function related to peristaltic movement, and that the *DjMHC-B*-expressing muscles may have a function related to body movement. These functional differences may be important in pharynx-cavity formation during regeneration. *DjMHC-B*-expressing muscles disappeared completely from the body-wall layer in the cavity-forming region, whereas *DjMHC-A*-expressing muscles were accumulated there.

To understand the evolution of muscles, it is also interesting to analyze composition and expression pattern of actin genes in planarians. Since it has been well documented







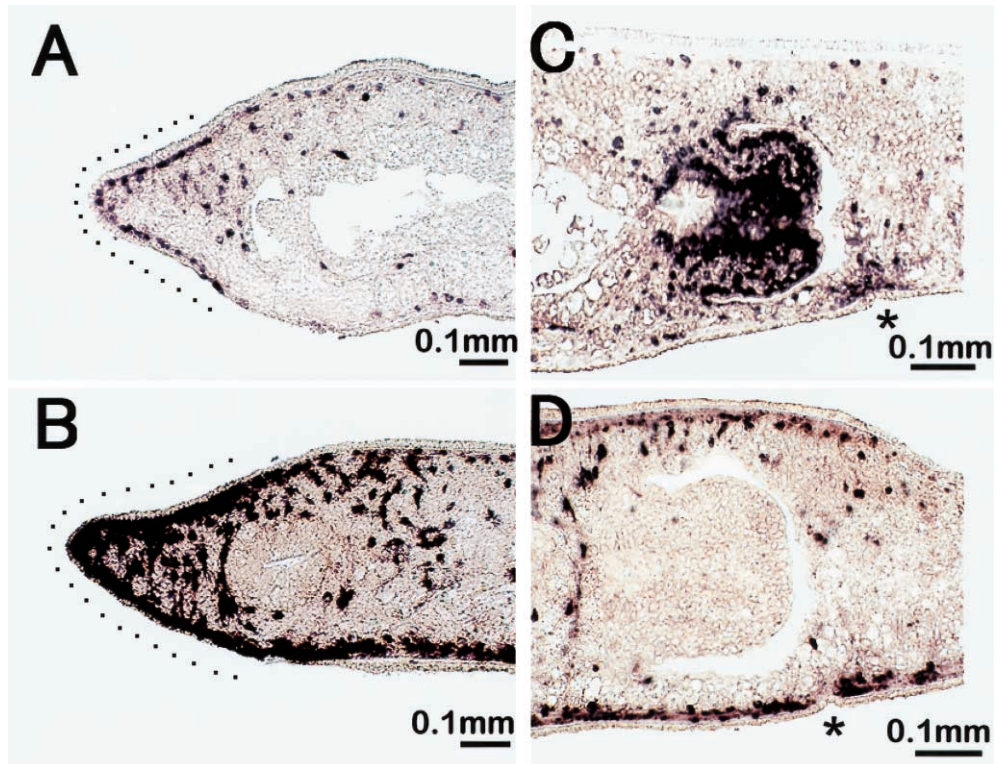
**Fig. 5.** Doubly stained *in situ* hybridization of sagittal sections using the *DjMHC-A* and *DjMHC-B* probes. Nuclei were counter-stained with Hoechst 33342 (blue). (A) A diagram of the position of sections. (B-D) Intestine and transverse muscle area. (B) Expression of *DjMHC-A* was observed in the muscles surrounding the intestinal duct (green). (C) Expression of *DjMHC-B* was observed in transverse muscles (red). (D) A double exposure of figures (B) and (C); *DjMHC-A* (green) and *DjMHC-B* (red). Dots indicate the intestinae. (E-G) Body-wall area. (E) Expression of *DjMHC-A* (green) (F) Expression of *DjMHC-B* (red). (G) A double exposure of figures (E) and (F); *DjMHC-A* (green) and *DjMHC-B* (red). I, intestinal duct. Scale bars, 50  $\mu\text{m}$ .

**Fig. 4.** Expression patterns of *DjMHC-A* and *DjMHC-B* in normal adult planarians. (A) Expression of *DjMHC-A*. (a) A whole-mount *in situ* hybridization. (b) A transverse section of the pharynx region. (c) A transverse section of the base of the pharynx. Arrowheads indicate expression in body-wall muscles. Arrows indicate expression in the muscles surrounding the intestinal ducts. Asterisks indicate expression in the cells in the mesenchymal space. (d) A high-magnification view of the pharynx region in a horizontal section. Arrows indicate expression in the pharynx-cavity muscle. Arrows indicate expression in the muscles surrounding the intestinal ducts. (B) Expression of *DjMHC-B*. (a-c) Arrows indicate expression in body-wall muscles. (a) A sagittal section. Arrowheads indicate transverse muscles. (b) A transverse section of the pharynx region. Arrowheads indicate transverse muscles. (c) A transverse section of the tail region. Short Arrows indicate horizontally oriented muscles. Open arrowheads indicate nucleus of the transverse muscles. Arrows indicate nucleus of the horizontally oriented muscles. (C) A diagram of the position of sections. I, intestinal duct. Scale bars, 0.5 mm (Aa) and 0.2 mm (Ab-d and Ba-c).

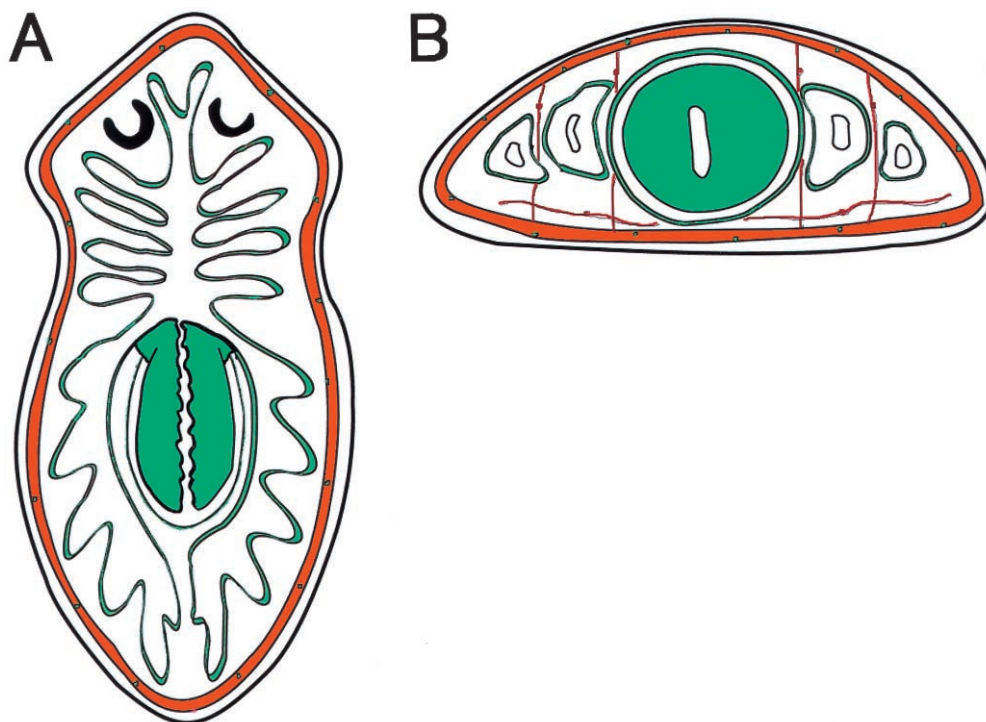
that the vertebrates have isoform-specific actin genes expressed in tissue- or developmental stage-specific manner (Vandekerckhove and Weber, 1979). Recently, such tissue- or developmental stage-specific expression patterns of actin genes of the invertebrates were also indicated in ascidians and arrow worms (Kusakabe *et al.*, 1991; Kusakabe *et al.*, 1995; Yasuda *et al.*, 1997). We need to describe expression pattern of actin genes in planarians and compare it with that of MHC genes to understand the characteristics and evolution of planarian muscles.

Cells expressing the *DjMHC-A* gene were observed in the mesenchymal region around the base of the pharynx, and they appear to participate in pharynx formation (Fig. 4A-c; asterisks). If they are pharynx-forming muscles, pharynx muscles may differentiate in the mesenchymal space and then migrate into the pharynx. We will need to clarify whether or not these





**Fig. 6.** Expression patterns of *DjMHC-A* and *DjMHC-B* during the process of regeneration from tail piece. (A, B) Sagittal sections 2 days after amputation. The blastema region is surrounded by dots. (A) Expression of *DjMHC-A*. (B) Expression of *DjMHC-B*. (C, D) Sagittal sections 5 days after amputation. Asterisks indicate the pharynx-cavity-forming region. (C) Expression of *DjMHC-A*. (D) Expression of *DjMHC-B*. Scale bars, 0.1 mm.



**Fig. 7.** Summary of the expression domains of *DjMHC-A* (green) and *DjMHC-B* (red). (A) A horizontal section. (B) A transverse section.

cells are pharynx-forming muscles, in order to understand the mechanism of pharynx regeneration in planarians.

Unexpectedly, we could not isolate genes other than the two MHC genes by the  $\lambda$ ZAP expression-cloning system in spite of using the antiserum against living planarians. All of the forty-seven positive clones contained cDNA fragments more than 5.7 kb in size. Surprisingly, sequence analyses revealed that most of the cDNA fragments had been fused to the *LacZ* gene out of frame. These results suggested that the MHC products may be translated from internal methionines in the 5' regions. When we changed the expression-cloning system from the  $\lambda$ ZAP system to the  $\lambda$ gt11 system, we succeeded in isolating a variety of genes with the same antiserum (data not shown). We therefore concluded that the characteristics of the  $\lambda$ ZAP system may be the reason why MHC genes were selectively isolated by immunoscreening using the antiserum against living planarians as a probe.

### ACKNOWLEDGMENTS

We thank Dr. Y. Umesono for encouragement. This work was supported by Special Coordination Funds for Promoting Science and Technology to K.A. and Grant-in-Aid for Scientific Research on Priority Areas to K.A. and K.W.

### REFERENCES

- Agata K, Soejima Y, Kato K, Kobayashi C, Umesono Y, Watanabe K (1998) Structure of the planarian central nervous system (CNS) revealed by neuronal cell markers. *Zool Sci* 15: 433–440
- Bueno D, Bagnà J, Romeo R (1997) Cell-, tissue-, and position-specific monoclonal antibodies against the planarian *Dugesia (Girardia) tigrina*. *Histochem Cell Biol* 107: 139–149
- Burr AH, Gans C (1998) Mechanical significance of obliquely striated architecture in nematode muscle. *Biol Bull* 194: 1–6
- Cebria F, Vispo M, Newmark P, Bueno D, Romero R (1997) Myocyte differentiation and body wall muscle regeneration in the planarian *Girardia tigrina*. *Dev Genes Evol* 207: 306–316
- Dibb NJ, Maruyama IN, Krause M, Karn J (1988) Sequence analysis of the Complete *Caenorhabditis elegans* myosin heavy chain gene family. *J Mol Biol* 205: 603–613
- Dowar BL (1973) A combined relaxing agent and fixative for triclads (planarians). *Stain Technol* 48: 96–94
- Epstein HF, Waterston RH, Brenner S (1974) A mutant affecting the heavy chain of myosin *Caenorhabditis elegans*. *J Mol Biol* 90: 291–300
- Garcea RL, Schachat F, Epstein HF (1978) Coordinate synthesis of two myosins in wild-type and mutant nematode muscle during larval development. *Cell* 15: 421–428
- Hanson J (1956) The structure of the smooth muscle fibres in the body wall of the earthworm. *J Biophysic Biochem Cytol* 3: 111–127
- Hori I (1983) Differentiation of myoblasts in the regenerating planarian *Dugesia japonica*. *Cell Differ* 12: 155–163
- Kusakabe T, Suzuki J, Saiga H, Jeffery WR, Makabe KW, Satoh N (1991) Temporal and spatial expression of a muscle actin gene during embryogenesis of the ascidean *Halocynthia roretzi*. *Dev Growth Differ* 33: 227–234
- Kusakabe T, Hikosaka A, Satoh N (1995) Coexpression and promoter function in two muscle actin gene complexes of different structural organization in the ascidean *Halocynthia roretzi*. *Dev Biol* 169: 461–472
- Lowy J, Hanson J (1962) Ultrastructure of invertebrate smooth muscle. *Physiol Rev* 42, Suppl 5: 34–47
- MacRae EK (1963) Observation on the fine structure of pharyngeal muscle in the planarian *Dugesia tigrina*. *J Cell Biol* 18:651–662
- Miller DM, Stockdale FE, Kern J (1986) Immunological identification of the genes encoding the four myosin heavy chain isoforms of *Caenorhabditis elegans*. *Proc Nat Acad Sci USA* 83: 2305–2309
- Morita M (1965) Electron microscopic studies on planaria. I. Fine structure of muscle fiber in the head of the planarian *Dugesia dorotocephala*. *J Ultrastruct Res* 13: 383–395
- Sarnat HB (1984) Muscle histochemistry of the planarian *Dugesia tigrina* (Turbellaria: Tricladida): implications in the evolution of muscle. *Trans Am Microsc Soc* 103: 284–294
- Schuchert P, Reber-Müller S, Schmid V (1993) Life stage specific expression of a myosin heavy chain in the hydrozoan *Podocoryne carnea*. *Differentiation* 51: 11–18
- Takahashi M, Kawamoto S, Adelstein RS (1992) Evidence for inserted sequences in the head region of non-muscle myosin specific to the nervous system. *J Biol Chem* 267: 17864–17871
- Umesono Y, Watanabe K, Agata K (1997) A planarian *orthopedia* homolog is specifically expressed in the branch region of both the mature and regenerating brain. *Dev Growth Differ* 39: 723–727
- Vandekerckhove J, Weber K (1979) The complete amino acid sequence of actins from bovine aorta, bovine heart, bovine fast skeletal muscle and rabbit slow skeletal muscle. *Differentiation* 14: 123–133
- Warrick HM, Spudich JA (1987) Myosin structure and function in cell motility. *Ann Rev Cell Biol* 3: 379–421
- Yasuda E, Goto T, Makabe KW, Satoh N (1997) Expression of actin genes in the arrow worm *Paraspadella gotoi* (Chaetognatha). *Zool Sci* 14: 953–960

(Received June 30, 1998 / Accepted August 6, 1998)