



---

## **Alternating Temperatures Affect the Performance of Trichogramma Species**

Authors: Firake, D. M., and Khan, M. A.

Source: Journal of Insect Science, 14(41) : 1-14

Published By: Entomological Society of America

URL: <https://doi.org/10.1673/031.014.41>



## Alternating temperatures affect the performance of *Trichogramma* species

D. M. Firake<sup>1a</sup> and M. A. Khan<sup>2</sup>

<sup>1</sup>Division of Entomology, ICAR Research Complex for NEH Region, Umiam, Meghalaya-793103, India

<sup>2</sup>Department of Entomology, G.B. Pant University of Agriculture and Technology, Pantnagar, Uttarakhand, India

### Abstract

The environmental compatibility of a biological control agent is an important aspect of successful reduction of agricultural pests. Temperature fluctuations during the day have a strong influence on the performance of laboratory-reared parasitoids. In field conditions, *Trichogramma* (Hymenoptera: Trichogrammatidae) wasps are exposed to variable temperatures during their development, which has a significant impact on their ability to control pest species. A simulation-based study was undertaken to evaluate the impact of variations in daily temperature on the pest-control abilities of female *Trichogramma* and their immature progenies. Considering the temperature variability of different agricultural zones of India, five temperature levels ranging from 20°C to 40°C were selected for daily short-term heat shocks to the immature progenies and egg-laying females of two major *Trichogramma* species. Intensity and frequency of thermal shocks showed inverse relationships with adult emergence, fecundity, and longevity of *T. chilonis* and *T. poliae*. Parasitoid pupae were found to be more tolerant to temperature variations than eggs and larvae. Fecundity and longevity of parasitoids were significantly reduced under high temperature shocks to egg-laying females. Sex ratio was significantly affected by high temperature shocks to the immature and adult stages. However, the effect was more severe in eggs. A female-biased sex ratio was apparent in both parasitoids throughout the experiment. Overall, daily short-term temperature shocks to different developmental stages of parasitoids showed radical effects on emergence, fecundity, longevity, and sex ratio of the progeny. Therefore, releases of parasitoids should be conducted when they are in their pupal stages during the morning and evening in order to achieve their highest effectiveness for pest management.

**Keywords:** biological attributes, egg parasitoids, temperature shocks, *Trichogramma chilonis*, *Trichogramma poliae*

**Correspondence:** a [dfirake@gmail.com](mailto:dfirake@gmail.com)

**Editor:** Michael Strand was editor of this paper.

**Received:** 23 May 2012 **Accepted:** 3 October 2012 **Published:** 15 March 2014

**Copyright:** This is an open access paper. We use the Creative Commons Attribution 3.0 license that permits unrestricted use, provided that the paper is properly attributed.

**ISSN:** 1536-2442 | Vol. 14, Number 41

**Cite this paper as:**

Firake DM, Khan MA. 2014. Alternating temperatures affect the performance of *Trichogramma* species. *Journal of Insect Science* 14:41. Available online: <http://www.insectscience.org/14.41>

## Introduction

*Trichogramma* (Hymenoptera: Trichogrammatidae) consists of over 600 species of egg parasitoids (Park et al. 2000), many of which are used in biological control programs against a wide range of lepidopterous pests (Corrigan and Laing 1994). *T. chilonis* Ishii is the most frequently encountered species in the Indian subcontinent and Southeast Asia (Nagarkatti and Nagaraja 1977), and *T. poliae* Nagaraja is a dominant parasitoid of the most destructive poplar defoliators, *Clostera cupreata* and *C. fulgurita*, in Himalayan regions of India (Ahmad et al. 1999, 2002). Both parasitoids are being deployed in mass in order to reduce lepidopteran pest populations in India.

Ecological compatibility of a biological control agent is an important aspect for successful agricultural pest control. The effectiveness of several bio-control attempts has been reduced due to adverse climatic conditions. Temperature is a key factor affecting the abundance, distribution, and parasitic capabilities of egg parasitoids (Bourchier and Smith 1996; Maceda et al. 2003). It has significant effects on developmental period and adult emergence of *Trichogramma* wasps (Foerster and Foerster 2009). Short-term exposure to 35°C temperature causes substantial mortality in *T. chilonis* (Nagarkati 1979). The functional response of the parasitoids is likely to change from one type to another with varying temperatures (Kfir 1983; Wang and Ferro 1998; Mohaghegh et al. 2001), resulting in altered foraging behavior (Zhang et al. 1983; Guo 1986). It has been reported that high temperature shocks on pupae and adults have significant impacts on fecundity, life duration, mobility, and efficacy of *Trichogramma* (Chihrane et al. 1993; 1997; Chihrane and Lauge 1994, 1996; Carriere and Boivin 1997).

Effects of temperature variation have been extensively studied on adult parasitoids, especially *Trichogramma* spp., but impacts on immature stages are often ignored. Additionally, several studies have been based on constant thermal exposure throughout the experimental period. However, subtropical countries like India, where the use of *Trichogramma* wasps as a biological control for agricultural pests is widely promoted, have variable temperatures during the day (usually six hours between 10:00 and 16:00). Regional temperature variability within the country ranges from less than 20°C in hilly regions to over 40°C in others. As a result of this variability, laboratory-reared parasitoids can be exposed to extreme temperature differences upon release, causing impacts that can affect their efficiency.

Simulation-based studies can give useful insight into the possible adverse effects of temperature variations on parasitoid performance. The present study was undertaken to evaluate the impacts of short-term exposure to extreme temperatures on the biological attributes of female *Trichogramma* and their immature progenies and to identify the best post-embryonic stage for their field release.

## Material and methods

### Study location

The experiment was carried out in the Biological Control Laboratory of the Department of Entomology, G.B. Pant University of Agriculture and Technology, Pantnagar, Uttarakhand, India, from 2006 to 2009.

### Source of insect cultures

Both parasitoids (*T. poliae* and *T. chilonis*) were cultured and maintained at the Biological Control Laboratory since 2004 in controlled conditions (25–27°C and 75 ± 5%

RH) on the eggs of rice moth, *Corcyra cephalonica* Stainton (Lepidoptera: Pyralidae).

### Maintenance of insect culture for the experiment

*C. cephalonica* was reared on a standard diet containing maize, *Zea mays* L. (Poales: Poaceae). Fresh eggs of the factitious host were collected regularly and used for both culture and experiments. Parasitoids were considered to be in the egg stage immediately after parasitization of host eggs. On the third day of parasitization (i.e., before blackening of host eggs), the parasitoid was considered to be in the larval stage. From blackening of the host eggs until adult emergence, the parasitoid was considered to be in the prepupal and pupal stages (Dahlan and Gordh 1996).

### Short-term exposure of immature stages of *Trichogramma* species to the variable temperatures

Paper strips containing 200 UV-light-treated eggs of *C. cephalonica* were exposed to large numbers of *T. poliae* and *T. chilonis* adults (about 500 females each) separately in glass test tubes (25 x 150 mm). Honey solution (10%) was provided as food to the adult parasitoids. After 24 hours of exposure, parasitized eggs were kept in an environmental chamber (OBROMAX, [www.obromax.com](http://www.obromax.com)) at  $20 \pm 1^\circ\text{C}$  temperature and  $75 \pm 5\%$  RH for the temperature shock period. After six hours of exposure, the parasitized eggs were kept at an ambient temperature ( $27 \pm 1^\circ\text{C}$ ) for the rest of the day. This process was repeated daily until adult emergence. A similar protocol was followed subsequently for different temperature shocks ( $25 \pm 1^\circ\text{C}$ ,  $30 \pm 1^\circ\text{C}$ ,  $35 \pm 1^\circ\text{C}$ , and  $40 \pm 1^\circ\text{C}$ ). A constant photoperiod (16:8 L:D) was maintained throughout the experiment, and exposure to temperature shocks were always conducted under light conditions.

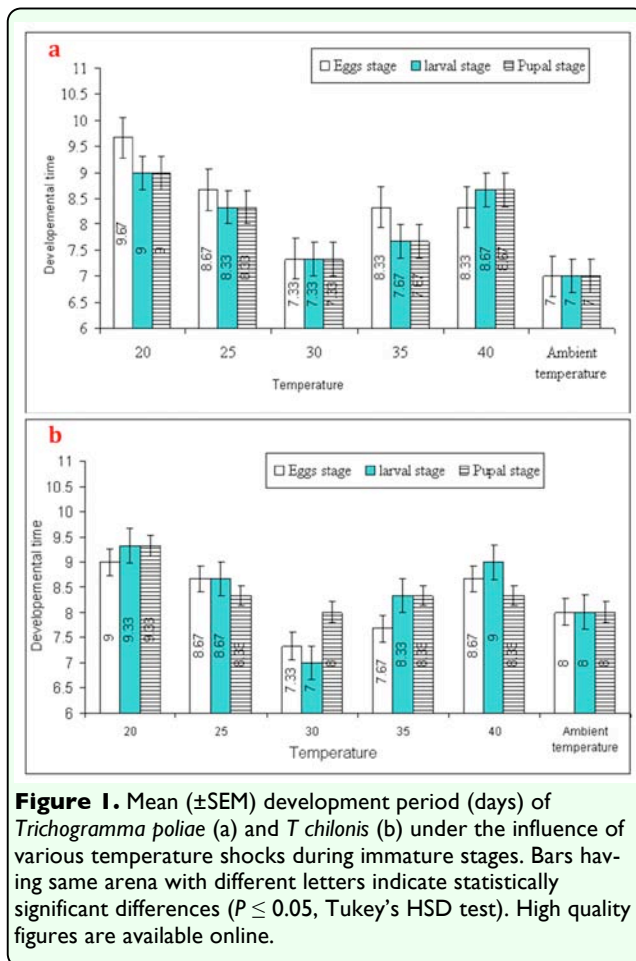
Continuous rearing of parasitoids at an ambient temperature ( $27 \pm 1^\circ\text{C}$ ) was used as an untreated control.

The different temperature shocks were given as explained above on the third day of parasitization (larval stage) and on fifth day of parasitization (pupal stage). Individuals (immature stages only) exposed to thermal treatments formed the  $G_0$  generation, and the  $G_1$  generation was composed of their progeny.

Immediately after adult emergence, 20 mated females from each replication were isolated by treatment in small glass vials (12 x 75 mm). For studies on fecundity of females ( $G_0$ ), 100 UV-treated fresh eggs of host *C. cephalonica* were provided for parasitization at ambient conditions ( $27 \pm 1^\circ\text{C}$  and  $75 \pm 5\%$  RH) until death. Honey solution (10%) was provided as food to the adult female parasitoids. Fresh host eggs were provided daily, and the strips containing parasitized host eggs were separated. After five to six days of parasitization, fecundity was calculated. Observations of female longevity ( $G_0$ ) were recorded each morning. Females in the progeny ( $G_1$ ) were separated on the basis of antennae under stereoscopic microscope and sex ratio was calculated.

### Short-term exposure of egg-laying *Trichogramma* species to variable temperatures

Both parasitoids were reared under ambient conditions up to pupal stage. After the adult emergence, 20 newly emerged and mated females were isolated in separate glass vials and 100 host eggs (UV-treated) were provided daily. Honey solution (10%) was provided as food for adult females. These vials were kept in an environmental chamber at  $20 \pm 1^\circ\text{C}$  temperature and  $75 \pm 5\%$  RH. After six hours of exposure to temperature shock, the vials



containing treated females were moved to ambient temperature ( $27 \pm 1^\circ \text{C}$ ) for the rest of the day. Parasitoids were exposed to different temperature shocks each day for six hours until death. The females exposed to variable temperatures formed the  $F_0$  generation and the  $F_1$  generation was composed of their subsequent progenies. Daily fecundity ( $F_0$ ), longevity ( $F_0$ ) and sex ratio of their progeny ( $F_1$ ) were recorded using the above procedure. The same protocol was followed subsequently for different temperature shocks ( $25 \pm 1^\circ \text{C}$ ,  $30 \pm 1^\circ \text{C}$ ,  $35 \pm 1^\circ \text{C}$ , and  $40 \pm 1^\circ \text{C}$ ). Parasitoids reared at an ambient temperature ( $27 \pm 1^\circ \text{C}$ ) without any temperature shocks were used as an untreated control.

## Experimental design and statistical analysis

The experiment on the effect of temperature shocks on immature stages of *Trichogramma* spp. was carried out with respect to two factors, temperature shock levels and life stage of parasitoids, with a completely randomized design. Data obtained in this study were analyzed using the general linear model (univariate analysis) in statistical software SPSS 14.0 (IBM, [www.ibm.com](http://www.ibm.com)). Data obtained during the experiment regarding temperature effects on gravid *Trichogramma* females were analyzed using one way ANOVA ( $P = 0.05$ ). Tukey's HSD test was applied to determine differences between means.

## Results

### Influence of variable temperature shocks on biological attributes of *T. poliae* and *T. chilonis*

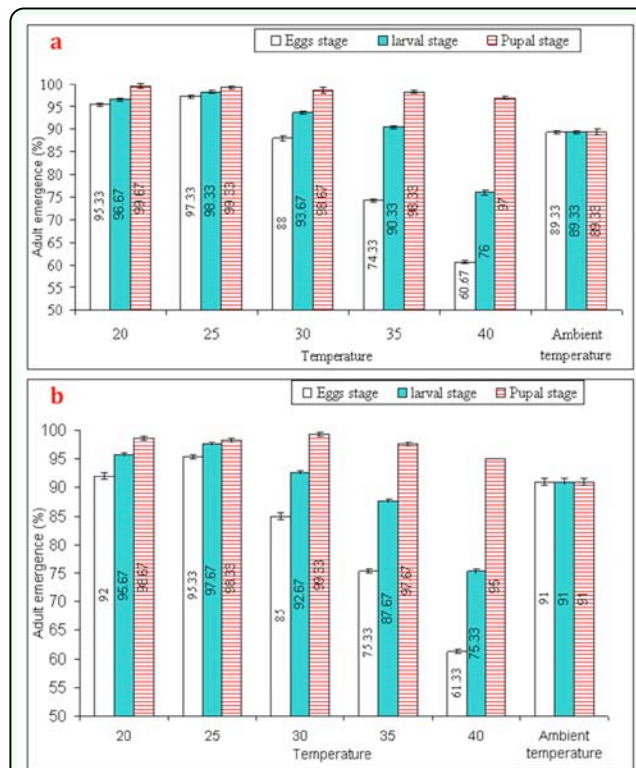
#### Developmental period of parasitoids ( $G_0$ ).

Significant variation in developmental period of *Trichogramma* species was observed as a result of experimental temperature shocks (Figure 1). Developmental period of *T. poliae* was remarkably extended ( $F = 9.15$ ,  $P < 0.01$ ,  $df = 5$ ) due to both low and high temperature shocks. A similar trend was found in *T. chilonis* ( $F = 5.78$ ,  $P < 0.01$ ,  $df = 5$ ). Differences in developmental period of *T. poliae* and *T. chilonis* were not significant when different stages of parasitoids were exposed to variable temperatures ( $F = 0.47$ ,  $P = 0.62$ ,  $df = 2$ , and  $F = 0.28$ ,  $P = 0.75$ ,  $df = 2$ , respectively).

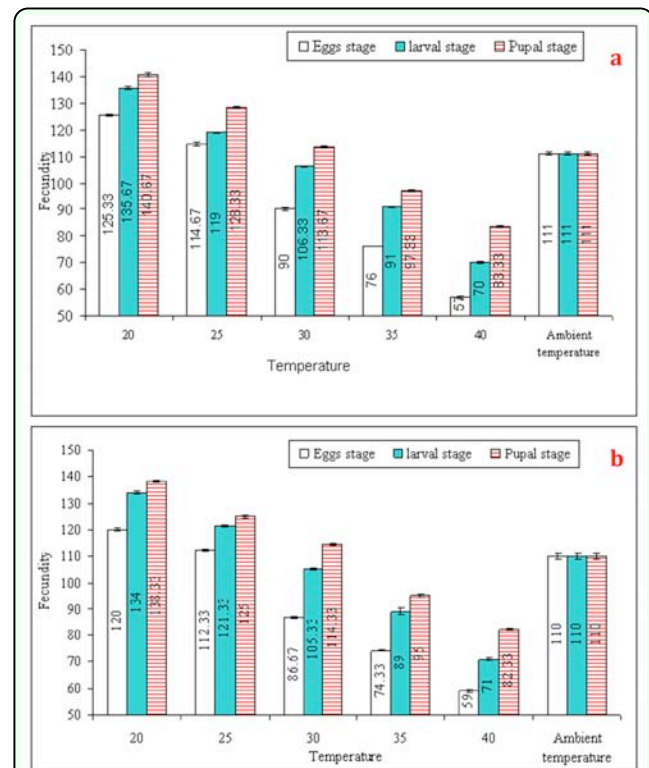
#### Adult emergence of parasitoids ( $G_0$ ).

Adult emergence of *T. poliae* and *T. chilonis* was also significantly affected ( $F = 1148.24$ ,  $P < 0.01$ ,  $df = 5$ , and  $F = 934.58$ ,  $P < 0.01$ ,  $df = 5$ , respectively) as a result of daily short-term exposure to higher temperatures (Figure 2). Furthermore, adult emergence was significant-





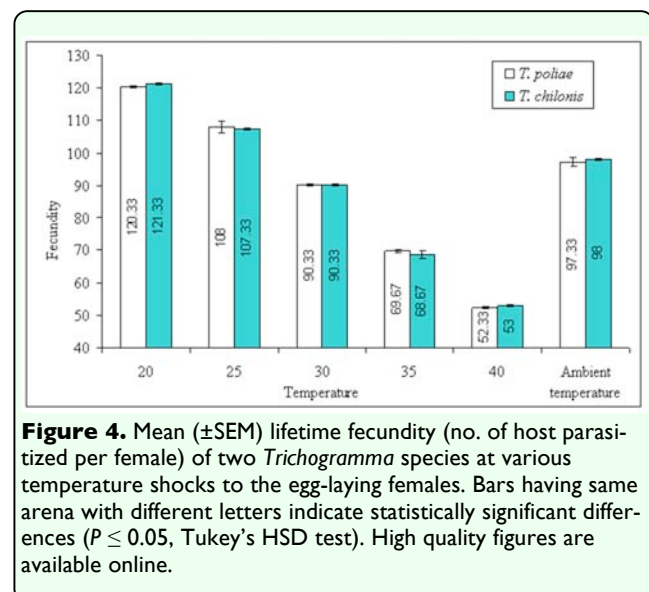
**Figure 2.** Mean ( $\pm$ SEM) adult emergence (%) of *Trichogramma poliae* (a) and *T. chilonis* (b) under the influence of various temperature shocks during immature stages. Bars having same arena with different letters indicate statistically significant differences ( $P < 0.05$ , Tukey's HSD test). High quality figures are available online.



**Figure 3.** Mean ( $\pm$ SEM) lifetime fecundity (no. of host parasitized per female) of *Trichogramma poliae* (a) and *T. chilonis* (b) under the influence of various temperature shocks during immature stages. Bars having same arena with different letters indicate statistically significant differences ( $P \leq 0.05$ , Tukey's HSD test). High quality figures are available online.

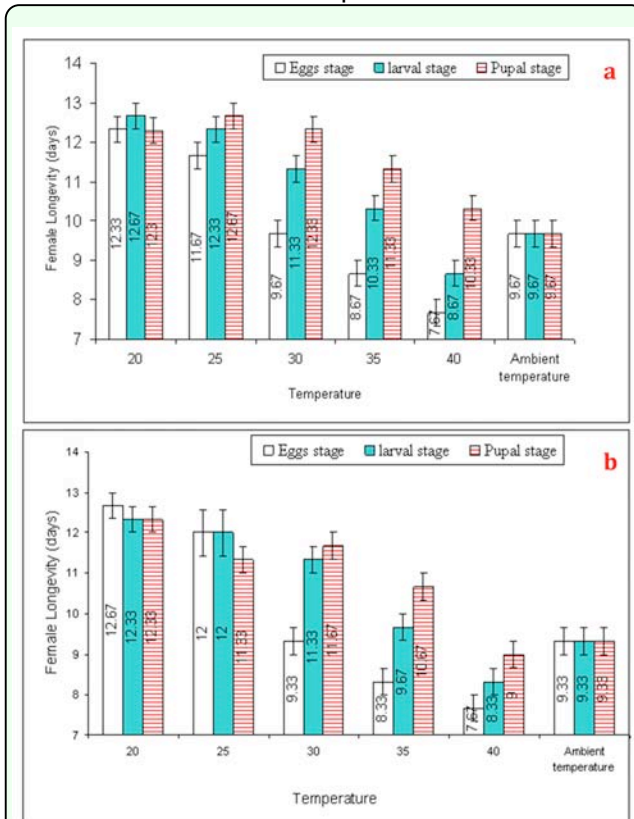
ly affected when egg-stage *T. poliae* ( $F = 1682.17$ ,  $P < 0.01$ ,  $df = 2$ ) and *T. chilonis* ( $F = 1600$ ,  $P < 0.01$ ,  $df = 2$ ) were exposed. Significant differences were also noticed between different temperature shocks and life stages of *T. poliae* ( $F = 360.77$ ,  $P < 0.01$ ,  $df = 10$ ) and *T. chilonis* ( $F = 251.51$ ,  $P < 0.01$ ,  $df = 10$ ).

**Fecundity of parasitoids ( $G_0$  and  $F_0$ ).** Fecundity of *T. poliae* ( $G_0$ ) and *T. chilonis* ( $G_0$ ) was significantly reduced ( $F = 2294$ ,  $P < 0.01$ ,  $df = 5$  and  $F = 3176.98$ ,  $P < 0.01$ ,  $df = 5$ , respectively) as a result of high temperature shocks (Figure 3). The effect was more severe when the *T. poliae* ( $F = 618.49$ ,  $P < 0.01$ ,  $df = 2$ ) and *T. chilonis* egg-stage ( $F = 996.61$ ,  $P < 0.01$ ,  $df = 2$ ) was exposed than when the larval and pupal stages were exposed. A significant reduction in fecundity was also observed



**Figure 4.** Mean ( $\pm$ SEM) lifetime fecundity (no. of host parasitized per female) of two *Trichogramma* species at various temperature shocks to the egg-laying females. Bars having same arena with different letters indicate statistically significant differences ( $P \leq 0.05$ , Tukey's HSD test). High quality figures are available online.

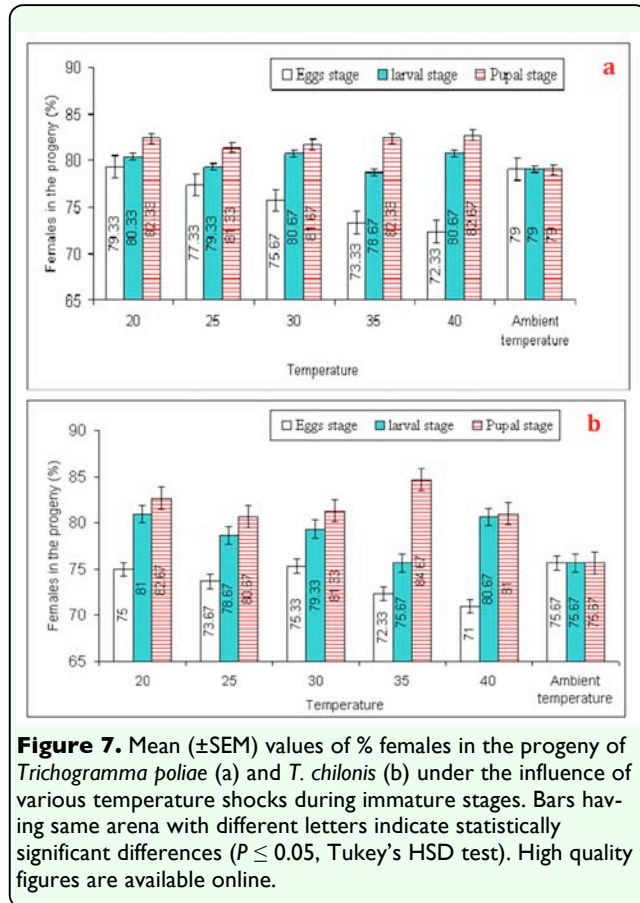
when egg-laying females ( $F_0$ ) of *T. poliae* ( $F = 675.98$ ,  $P < 0.01$ ,  $df = 5$ ) and *T. chilonis* ( $F = 796.22$ ,  $P < 0.01$ ,  $df = 5$ ) were exposed to various temperature shocks (Figure 4).



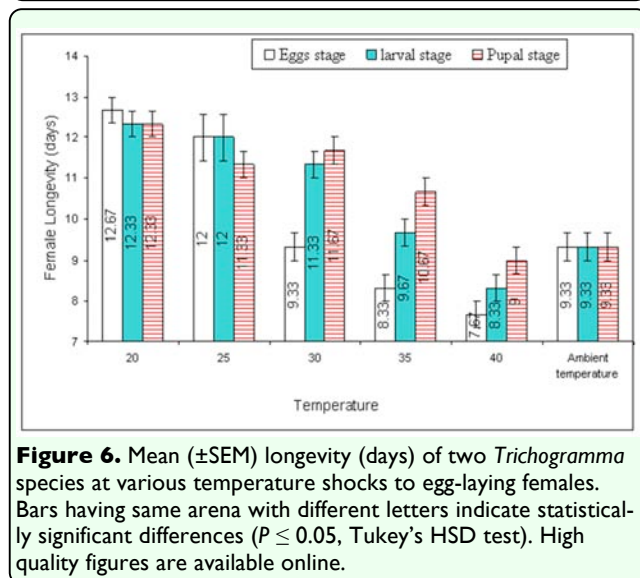
**Figure 5.** Mean ( $\pm$ SEM) longevity (days) of *Trichogramma poliae* (a) and *T. chilonis* (b) under the influence of various temperature shocks during immature stages. Bars having same arena with different letters indicate statistically significant differences ( $P \leq 0.05$ , Tukey's HSD test). High quality figures are available online.

### Female longevity of parasitoids ( $G_0$ and $F_0$ ).

The life spans of female *T. poliae* ( $G_0$ ) and *T. chilonis* ( $G_0$ ) were considerably reduced ( $F = 53.73$ ,  $P < 0.01$ ,  $df = 5$ . and  $F = 49.97$ ,  $P < 0.01$ ,  $df = 5$ , respectively) from short-term exposure to 40°C temperature shocks (Figure 5). Longevity of female *T. poliae* ( $G_0$ ) and *T. chilonis* ( $G_0$ ) was higher when exposed to higher temperatures at the pupal stage ( $F = 11.40$ ,  $P < 0.01$ ,  $df = 2$ , and  $F = 7.54$ ,  $P < 0.01$ ,  $df = 2$ , respectively) than the larval and egg stage. Increased intensity and frequency of thermal shocks caused significant effects on longevity of *T. poliae* ( $F_0$ ) ( $F = 16.89$ ,  $P < 0.01$ ,  $df = 5$ ) and *T. chilonis* ( $F_0$ ) ( $F = 53.40$ ,  $P < 0.01$ ,  $df = 5$ ) (Figure 6).



**Figure 7.** Mean ( $\pm$ SEM) values of % females in the progeny of *Trichogramma poliae* (a) and *T. chilonis* (b) under the influence of various temperature shocks during immature stages. Bars having same arena with different letters indicate statistically significant differences ( $P \leq 0.05$ , Tukey's HSD test). High quality figures are available online.



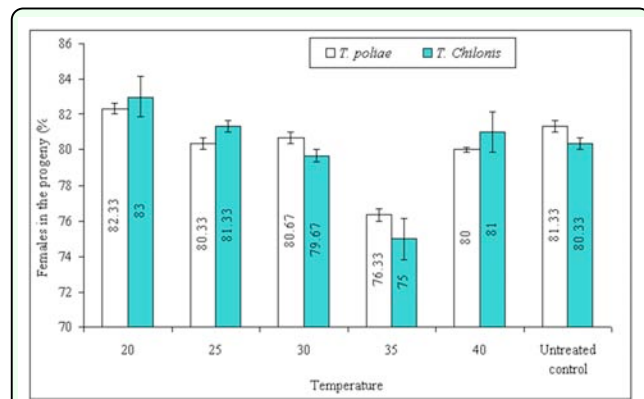
**Figure 6.** Mean ( $\pm$ SEM) longevity (days) of two *Trichogramma* species at various temperature shocks to egg-laying females. Bars having same arena with different letters indicate statistically significant differences ( $P \leq 0.05$ , Tukey's HSD test). High quality figures are available online.

**Sex ratio of the progeny ( $G_1$  and  $F_1$ ).** No significant differences were noticed in sex ratio of the progeny ( $G_1$ ) of *T. poliae* ( $F = 0.50$ ,  $P = 0.77$ ,  $df = 5$ ) or *T. chilonis* ( $F = 0.60$ ,  $df = 5$ ); when their mothers ( $G_0$ ) were exposed to various temperature shocks (Figure 7). However, the sex ratio of the progeny of *T.*

*poliae* and *T. chilonis* ( $G_1$ ) decreased considerably ( $F = 9.93$ ,  $P < 0.01$ ,  $df = 2$ , and  $F = 11.38$ ,  $p < 0.01$ ,  $df = 2$ , respectively) under high temperature shocks at the egg stage, favoring females (Figure 7). Furthermore, the sex ratio of *T. poliae* ( $F_1$ ) was significantly affected ( $F = 13.37$ ,  $P < 0.01$ ,  $df = 5$ ) as a result of various temperature shocks to egg-laying females ( $F_0$ ) (Figure 8). A parallel trend was observed in *T. chilonis* ( $F_1$ ) ( $F = 10.24$ ,  $P < 0.01$ ,  $df = 5$ ) (Figure 8). Interestingly, sex ratio of both the parasitoid species was female-biased under all temperature shock levels.

## Discussion

Temperature is the most important variable impacting the development of invertebrates. The developmental period of insects largely depends on environmental temperatures. In this study, both high and low temperature shocks extended the developmental period of *Trichogramma* species. Changes in enzyme levels, the properties of the cell membrane, as well as other physio-chemical changes induced by fluctuating temperatures may have extended the developmental period of insects (Wang et al. 2004). The relationship between temperature and development is often linear throughout the range of temperatures suitable for insect development and becomes sigmoid throughout the temperature ranges unsuitable for insect development (Andrewartha and Birch 1954; Lactin et al. 1995; Arbab et al., 2006). Therefore, the results of our study are in agreement with several reports on other *Trichogramma* species (Butler and Lopez 1980; Calvin et al. 1984; Naranjo 1993; Park et al. 2000; Suverkropp et al. 2001). Extended developmental period under fluctuating temperatures may also be attributed to reduced metabolic conditions for prolonged periods (Hassan et al. 1990; Tezze and Botto 2004).



**Figure 8.** Mean ( $\pm$ SEM) values of % females in the progeny of two *Trichogramma* species at various temperature shocks to egg-laying females. Bars having same arena with different letters indicate statistically significant differences ( $P \leq 0.05$ , Tukey's HSD test). High quality figures are available online.

Developmental period was least affected when parasitoids were exposed in the pupal stage. This stage has been suggested to be more tolerant of temperature extremes. Interestingly, the developmental period of *T. poliae* was relatively less affected, even under higher temperatures, possibly a result of originating from a warmer climate (Nagaraja 1973).

When temperature deviates from its optimum, mortality rate increases and premature mortality is observed among individuals (Chihrane and Lauge 1996). This inverse relationship of temperature with emergence of *Trichogramma* has been supported by several studies (Ram and Sharma 1977; Lopez and Morrison 1980; Chihrane et al. 1993; Miura and Kobayashi 1993; Park et al. 2000). Reduced *Trichogramma* emergence has also been reported even at 32°C (Consoli and Parra 1995), 35°C (Harrison et al. 1985; Scholler and Hassan 2001; Ayvaz et al. 2008), and 42°C (Fye and Larsen 1969). Lower emergence at higher temperatures might be due to higher mortality of parasitoids inside the host egg itself. Under such conditions, parasitoids may be unable to perforate the chorion of host egg, ultimately blocking emergence (Chihrane and Lauge 1996).



In this study, adult emergence was more significantly affected when parasitoids were exposed in the egg-stage, as opposed to beginning exposure at the larval and pupal stages. Desiccation of host eggs, prolonged heat stress, and sudden changes in metabolic activities might have affected parasitoid development, which ultimately reduced adult emergence. Insect pupae are usually comparatively harder than eggs and larvae; therefore, temperature-induced effects on the parasitoid pupae were less significant, even up to  $40 \pm 1^\circ\text{C}$ . Pupal stages in this study were given fewer thermal shocks than the other life stages. However, extreme temperature exposure of pupal-stage *T. pretiosum* (Lopez and Morrison 1980) and *T. brassicae* (Chihrane and Lauge 1996) significantly reduced the emergence of adults (Hassan et al. 1988; Steidle et al. 2001; Mansfield and Mills 2002). This may be due to their originating from a colder climate. The parasitoids used in this study originated from a tropical climate.

Short-term exposure to high temperatures also had significant impacts on field parasitism of crop pests by *T. ostriniae* (Wang et al. 1997). A negative correlation between temperature and fecundity has been repeatedly reported in many parasitoids (Sorokina 1978; Zaslavskiy and May Phu Qui 1982; Atamirzaeva 1991; Pavlik 1992; Scott et al. 1997; Wang and Ferro 1998). In our findings, fecundity of parasitoids was considerably reduced due to high temperature shocks, in congruence with earlier reports on *Trichogramma* species (Calvin et al. 1984; Chihrane et al. 1993; Chihrane and Lauge 1996; Reznik et al. 1997; Maisonhaute et al. 1999; Singh and Ram 2006; Reznik and Vaghina 2006a, b; Ayvaz et al. 2008). Reduced fecundity is linked to the disruption of ovarian function as a result of high temperatures during post-embryonic development. Possibilities of some pathological

effects have also been reported (partial male sterility and female oogenesis suppression) in *T. brassicae* due to heat shocks and their ultimate influence on oviposition (Chihrane et al. 1993; Chihrane and Laugé 1994, 1996). In this study, the pupal stage was found to be relatively tolerant, which could be due to their hardiness and fewer thermal shocks, as previously described.

Longevity was also significantly reduced due to high temperature shocks given during immature and adult stages. However, life span was comparatively longer when parasitoids were exposed in their pupal stages. Shortened longevity could be due to shrinkage or desiccation of host eggs, which ultimately result in the formation of weak parasitoids. Usually, malnourished parasitoids (food-deprived females) live shorter life spans than well-fed parasitoids (Reznik et al. 1997, 2003). In this investigation, longevity was inversely related to temperature shocks, suggesting that temperature plays a significant role on overall parasitization. Therefore, the results are in accordance with several reports on other *T.* species (Lund 1938; Ram and Sharma 1977; Pak and Oatman 1982; Harrison et al. 1985; Rogor 1988; Chihrane et al. 1991; Bouchier and Smith 1996; Babi and Nabham 1998; Ayvaz et al. 2008).

Thermal fluctuations given during immature stages ( $G_0$ ) had lesser effects on the sex ratio of the progeny ( $G_1$ ) and less effects on the pupal stage compared to egg and larval stages. Tolerance of *T. chilonis* pupae to extreme temperatures has been reported (Singh and Ram 2006), whereas sex ratio was relatively unchanged due to temperature variations. However, the sex ratio of the progeny of both parasitoids ( $F_1$ ) was significantly affected due to thermal shocks in this study. The effects on the sex ratio of the progeny were determined

by the rearing temperature during the mother's development (Bower and Stern 1996). In our study, the mothers ( $F_0$ ) were reared at ambient conditions and exposed to variable temperatures during the adult stage only. This might have affected the sex ratio of the progeny ( $F_1$ ). A similar relationship between temperature and sex ratio has been observed in several *Trichogramma* species (Parker and Pinnell 1971; Harrison et al. 1985; Consoli and Parra 1995). In our experiment, a female-biased sex ratio consistently resulted when the mother was exposed to different temperature shocks, a trend that has been as reported in many trichogrammatids (Harrison et al. 1985; Haile et al. 2002; Saljoqi and YuRong 2004; Foerster and Foerster 2009).

### Conclusion

The results of our study suggest that high and low temperature shocks have considerable impacts on female parasitoid longevity, fecundity, adult emergence, and sex ratio of their progeny. Although the parasitoids in this study were found to survive and reproduce at temperatures up to 40°C, their releases for pest control would be more effective between  $23 \pm 1^\circ\text{C}$  and  $26 \pm 1^\circ\text{C}$ . Parasitoids should be released just before their emergence (during the pupal stage) and preferably during the morning or evening in order to achieve the highest level of effectiveness for better pest management.

### Acknowledgements

We are highly thankful to the Department of Biotechnology, Government of India, New Delhi, and the Dean of the College of Agriculture, G.B.P.U.A.&T., Pantnagar, India, for providing necessary laboratory facilities.

### References

- Ahmad M, Ahmad MJ, Mishra RK, Khan HR. 2002. Egg parasitoid, *Trichogramma chilonis* and its host range in India. *Indian Forester* 128: 412–420.
- Ahmad M, Jamal Ahmad MD, Mishra RK. 1997. Indigenous microhymenoptera parasitoid species associated with poplar defoliator, *Clostera cupreata* (Lepidoptera: Notodontidae). *Annals of Forestry* 5: 26–34.
- Ahmad M, Jamal Ahmad MD, Mishra RK. 1999. *Trichogramma poliae* Nagaraja (Hymenoptera: Trichogrammatidae): A new record on poplar defoliator, *Clostera cupreata* (Lepidoptera: Notodontidae). *Annals of Forestry* 7: 9–19.
- Andrewartha HG, Birch LC. 1954. *The Distribution and Abundance of Animals*. Chicago University Press.
- Arbab A, Kontodimas D, Sahragard A. 2006. Estimating development of *Aphis pomi* (De Geer) (Homoptera: Aphididae) using linear and nonlinear models. *Environmental Entomology* 35: 1208–1215.
- Atamirzaeva TM. 1991. Biological characteristics of *Trichogramma sugonjaevi* (Hymenoptera: Trichogrammatidae) under laboratory conditions. (pp. 5–6) In: *Trichogramma (Biology, Development, Application)*. *Proceedings of the Third All-Union Conference on Trichogramma*.
- Ayvaz A, Karasu E, Karaborklu J, Tuncbilek AS. 2008. Effects of cold storage, rearing temperature, parasitoid age and irradiation on the performance of *Trichogramma evanescens* Westwood (Hymenoptera:

Trichogrammatidae). *Journal of Stored Product Research* 44: 232–240.

Babi A, Nabham MA. 1998. Effect of temperature on some biological characteristics of the Syrian populations of *Trichogramma principium* Sugonyaev and Sorokina (Hymenoptera: Trichogrammatidae). *Entomophaga* 29: 369–379.

Bourchier RS, Smith SM. 1996. Influence of environmental conditions and parasitoid quality on field performance of *Trichogramma minutum*. *Entomologia Experimentalis et Applicata* 80: 461–468.

Butler GD, Lopez JD. 1980. *Trichogramma pretiosum* development in two hosts in relation to constant and fluctuating temperatures. *Annals of Entomological Society of America* 73: 671–673.

Calvin DD, Knapp MC, Welch SM, Poston FL, Elzinga RJ. 1984. Impact of environmental factors on *Trichogramma pretiosum* reared on southwestern corn borer eggs. *Environmental Entomology* 13: 774–780.

Carriere Y, Boivin G. 1997. Evolution of thermal sensitivity of parasitization capacity in egg parasitoids. *Evolution* 51: 2028–2032.

Chihrane J, Derrien A, Laugé G. 1997. Locomotor activity of *Trichogramma brassicae* (Hymenoptera) under the influence of high temperature shocks. *Journal of Insect Behavior* 10: 203–211.

Chihrane J, Lauge G. 1994. Incidences de chocs de températures élevées sur la lignée germinale male de *Trichogramma brassicae* (Hymenoptera: Trichogrammatidae). *Entomophaga* 39: 11–20.

Chihrane J, Lauge G. 1996. Loss of parasitization efficiency of *Trichogramma brassicae* (Hymenoptera: Trichogrammatidae) under high temperature conditions. *Biological Control* 87: 85–89.

Chihrane J, Lauge G, Hawtlzky N. 1993. Effect of high temperature shocks on the development and biology of *Trichogramma brassicae*. *Entomophaga* 38: 185–192.

Chihrane J, Lauge G, Hawtlzky N, Durier C, Barry P. 1991. Effect of high temperature shocks applied during pupal instar on characteristics of *Trichogramma brassicae*. *Les Colloques de l'INRA* 56: 63–64.

Consoli FL, Parra JRP. 1995. Effects of constant and alternating temperatures on *Trichogramma galloi* Zucchi (Hymenoptera: Trichogrammatidae) biology. I. Development and thermal requirements. *Journal of Applied Entomology* 119: 415–418.

Corrigan JE, Laing JE. 1994. Effects of the rearing host species and the host species attacked on performance by *Trichogramma minutum* Riley (Hymenoptera: Trichogrammatidae). *Biological Control* 23: 755–760.

Dahlan AN, Gordh G. 1996. Development of *Trichogramma australicum* Girault (Hymenoptera : Trichogrammatidae) on *Helicoverpa armigera* (Hubner) eggs (Lepidoptera: Noctuidae). *Australian Journal of Entomology* 35: 337–344.

Foerster MR, Foerster LA. 2009. Effects of temperature on the immature development and emergence of five species of *Trichogramma*. *BioControl* 54: 445–450.

- Fye RE, Larsen DJ. 1969. Preliminary evaluation of *Trichogramma minutum* as a released regulator of Lepidopterous pests of cotton. *Journal of Economic Entomology* 62: 1291–1296.
- Guo X. 1986. Bionomics of *Trichogramma ostrinia* Pang et Chen. *Chinese Journal of Biological Control* 2: 148–152.
- Haile AT, Hassan SA, Ogot CKPO, Baumgartner J, Sithanatham S, Monje JC, Zebitz CPW. 2002. Temperature dependent development of four egg parasitoid *Trichogramma* species (Hymenoptera: Trichogrammatidae). *Biocontrol Science and Technology* 12: 555–567.
- Harrison WW, King EG, Ouzts JD. 1985. Development of *Trichogramma exiguum* and *T. pretiosum* at five temperature regimes. *Environmental Entomology* 14: 118–121.
- Hassan SA, Kohler E, Rost WM. 1988. Mass production and utilization of *Trichogramma*: Control of the codling moth *Cydia pomonella* and the summer fruit tortrix moth *Adoxophyes orana* (Lepidoptera: Tortricidae). *Entomophaga* 33: 413–420.
- Honek A, Kocourek F. 1990. Temperature and development time in insects: A general relationship between thermal constants. *Zoologische Jahrbuecher Systematik* 117: 401–439.
- Ionascu G. 1992. *Effets de temperatures e'leve'es lors du re'veil de la pre'nympe de Trichogramma brassicae; ses conse'quences sur la fin du de'veloppement pre'imaginal et la biologie des adultes*. Me'moire D.E.A., University of Paris XI.
- Kfir R. 1983. Functional response to host density by the egg parasite *Trichogramma pretiosum*. *Entomophaga* 28: 345–353.
- Lactin DJ, Holliday NJ, Johnson DL, Craigen R. 1995. Improved rate model of temperature dependent development by arthropods. *Environmental Entomology* 24: 68–75.
- Lopez JD, Morrison RK. 1980. Effects of high temperature on *Trichogramma pretiosum* programmed for field release. *Journal of Economic Entomology* 73: 667–670.
- Lund HO. 1938. Studies on longevity and productivity in *Trichogramma evanescence*. *Journal of Agricultural Research* 56: 421–439.
- Maceda A, Homhmann CL, Honorio RDS. 2003. Temperature effects of *Trichogramma pretiosum* Riley and *Trichogrammatoidae annulata* De Santis. *Journal of Brazilian Archives of Biology and Technology* 46: 27–32.
- Maisonhaute C, Chihrane J, Lauge G. 1999. Induction of thermo-tolerance in *Trichogramma brassicae* (Hymenoptera: Trichogrammatidae). *Environmental Entomology* 28: 116–122.
- Mansfield S, Mills NJ. 2002. Host egg characteristics, physiological host range, and parasitism following inundative releases of *Trichogramma platneri* (Hymenoptera, Trichogrammatidae) in walnut orchards. *Environmental Entomology* 31: 723–731.
- Miura K, Kobayashi M. 1993. Effects of temperature on the development of *Trichogramma chilonis* Ishii (Hymenoptera: Trichogrammatidae), an egg parasitoid of the



diamond back moth. *Applied Entomology and Zoology* 28: 393–396.

Mohaghegh J, Clercq De P, Tirry L. 2001. Functional response of the predators *Podisus maculiventris* (Say) and *P. nigrispinus* (Dallas) (Heteroptera: Pentatomidae) to the beet army worm, *Spodoptera exigua* (Hubner) (Lepidoptera: Noctuidae): Effect of temperature. *Journal of Applied Entomology* 125: 131–134.

Nagaraja H. 1973. On some new species of Indian *Trichogramma* (Hymenoptera: Trichogrammatidae). *Orient insects* 7: 275–278

Nagarkatti S. 1979. Experimental comparison of laboratory reared vs. wild type *Trichogramma chilonis* (Hymenoptera: Trichogrammatidae). II. Tolerance of non-optimal temperatures. *Entomophaga* 24: 417–421.

Nagarkatti S, Nagaraja H. 1977. Biosystematics of *Trichogramma* and *Trichogrammatoidea* species. *Annual Review of Entomology* 22: 157–176.

Naranjo SE. 1993. Life history of *Trichogrammatoidea bactrae* (Hymenoptera: Trichogrammatidae), an egg parasitoid of pink bollworm (Lepidoptera: Gelechiidae), with emphasis on performance of high temperature. *Environmental Entomology* 22: 1051–1059.

Park YK, Lee HP, Lee KS. 2000. Effect of temperature on the biology of *Trichogramma dendrolimi* (Hymenoptera: Trichogrammatidae) reared on a factitious host, *Antheraea pernyi* (Lepidoptera: Saturniidae) egg. *Journal of Asia-Pacific Entomology* 3: 65–70.

Pavlik J. 1992. The effect of temperature on parasitization activity in *Trichogramma* spp. (Hymenoptera, Trichogrammatidae). *Zoologische Jahrbucher Abteilung fur allgemeine Zoologie und Physiologie der Tiere* 96: 417–425.

Ram A, Sharma AK. 1977. Laboratory assessment at different combinations of temperature and humidity on the efficiency of a strain of *Trichogramma fassiatum* (Perkins) (Hymenoptera: Trichogrammatidae) selected for high fecundity. *Journal of Entomological Research* 1: 47–54.

Ramesh B, Baskaran P. 1996. Development and response of four species of *Trichogramma* to heat shocks. *Entomophaga* 41: 267–277.

Reznik SY, Vaghina NP. 2006a. Temperature effects on induction of parasitization by females of *Trichogramma principium* (Hymenoptera: Trichogrammatidae). *Entomological Review* 86: 133–138.

Reznik SY, Vaghina NP. 2006b. Heat shock influences on parasitization of the angoumois grain moth *Sitotroga cerealella* Oliv. (Lepidoptera: Gelechiidae) eggs by the egg parasitoid *Trichogramma principium* Sugonyaev and Sorokina (Hymenoptera: Trichogrammatidae) females. *Entomological Review* 86: 861–865.

Reznik SY, Umarova TY, Voinovich ND. 1997. The influence of previous host age on current host acceptance in *Trichogramma*. *Entomologia Experimentalis et Applicata* 82: 153–157.

Reznik SY, Umarova TY, Voinovich ND. 2003. Egg retention in *Trichogramma* (Hymenoptera: Trichogrammatidae): Learning

or diapause? *Acta Societas Zoologicae Bohemicae* 67: 25–33.

Roger F. 1988. Effets de chocs de températures appliqués au stade nymphal de *Trichogramma maidis* (Hymenoptera: Trichogrammatidae) sur la biologie des adultes. *Memoire presente a E.N.I.T.A. Bordeaux*.

Russo J, Voegelé J. 1982. Influence de la température sur quatre espèces de trichogrammes (Hymenoptera: Trichogrammatidae) parasites de la pyrale dumais, *Ostrinia nubilalis* Hubn. (Lepidoptera: Pyralidae), I. Développement préimaginal. *Agronomie* 2: 509–516.

Russo J, Voegelé J. 1982a. Influence de la température sur quatre espèces de trichogrammes (Hymenoptera: Trichogrammatidae) parasites de la pyrale du maïs, *Ostrinia nubilalis* Hubn. (Lepidoptera: Pyralidae). II. Reproduction et survie. *Agronomie* 2: 517–524.

Saljoqi AUR, YuRong H. 2004. Effect of temperature on the development of *Trichogramma ostriniae* (Hymenoptera: Trichogrammatidae). *Journal of South China Agricultural University* 25: 43–46.

Scholler M, Hassan SA. 2001. Comparative biology and life tables of *Trichogramma evanescens* and *T. cacoeciae* with *Ephestia cautella* as host at four constant temperatures. *Entomologia Experimentalis et Applicata* 98: 35–40.

Scott M, Berrigan D, Hoffmann AA. 1997. Costs and benefits of acclimatization to elevated temperature in *Trichogramma carverae*. *Entomologia Experimentalis et Applicata* 85: 211–219.

Singh R, Ram P. 2006. Effect of high temperature shocks on performance of *Trichogramma chilonis* Ishii (Hymenoptera: Trichogrammatidae). *Journal of Biological Control* 20: 127–134.

Sorokina AP. 1978. New data on taxonomy and biology of the genus *Trichogramma* (Hymenoptera: Chalcidoidea). *Zoologicheskyy Zhurnal* 57: 1442–1444.

SPSS Inc. 2004. *SPSS for Windows, Release 13.0*. SPSS, Inc.

Steidle JLM, Rees D, Wright EJ. 2001. Assessment of Australian *Trichogramma* species (Hymenoptera: Trichogrammatidae) as control agents of stored product moths. *Journal of Stored Products Research* 37: 263–275.

Suverkropp BP, Bigler F, Van Lenteran JC. 2001. Temperature influences walking speed and walking activity of *Trichogramma brassicae* (Hymenoptera: Trichogrammatidae). *Journal of Applied Entomology* 125: 303–307.

Tezze AA, Botto EN. 2004. Effect of cold storage on the quality of *Trichogramma nerudai* (Hymenoptera: Trichogrammatidae). *Biological Control* 30: 11–16.

Wang B, Ferro DN. 1998. Functional responses of *Trichogramma ostriniae* (Hymenoptera: Trichogrammatidae) to *Ostrinia nubilalis* (Lepidoptera: Pyralidae) under laboratory and field conditions. *Environmental Entomology* 27: 752–758.

Wang B, Ferro DN, Hosmer DW. 1997. Importance of plant size, distribution of egg masses, and weather conditions on egg parasitism of the European corn borer,

*Ostrinia nubilalis* by *Trichogramma ostriniae* in sweet corn. *Entomologia Experimentalis et Applicata* 83: 337–345.

Wang B, Ferro DN, Wu J, Wang S. 2004. Temperature-dependent development and oviposition behavior of *Trichogramma ostriniae* (Hymenoptera: Trichogrammatidae), a potential biological control agent for the European corn borer (Lepidoptera: Crambidae). *Physiological Entomology* 33:777–793.

Zaslavskiy VA, May Phu Q. 1982. An experimental study of some factors affecting fecundity in *Trichogramma* Westwood (Hymenoptera: Trichogrammatidae). *Entomologicheskoe Obozrenie* 61: 724–736.

Zhang J, Wang J, Liu G, Yan Y. 1983. Influences of the humidities and temperature-humidity combinations on *Trichogramma ostriniae* Pang et Chen (Hymenoptera: Trichogrammatidae). *Natural Enemies of Insects* 5: 129–134.